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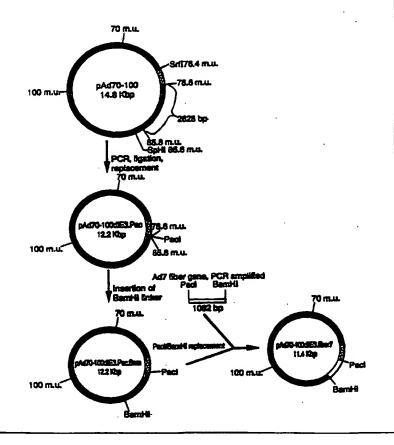
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(54) Title: CHIMERIC ADENOVIRAL COAT PROTEIN AND METHODS OF USING SAME

(57) Abstract

The present invention provides a chimeric adenoviral coat protein (particularly a chimeric adenovirus hexon protein). The chimeric adenovirus coat protein has a decreased ability or inability to be recognized by a neutralizing antibody directed against the corresponding wild-type adenovirus coat protein.



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CHIMERIC ADENOVIRAL COAT PROTEIN AND METHODS OF USING SAME

TECHNICAL FIELD OF THE INVENTION

The present invention relates to a chimeric adenoviral coat protein and a recombinant adenovirus comprising same. In particular, the invention provides a chimeric adenoviral hexon protein and a recombinant adenovirus comprising the chimeric adenoviral hexon protein. Such a recombinant adenovirus can be employed inter alia in gene therapy.

BACKGROUND OF THE INVENTION

In vivo gene therapy is a strategy in which nucleic acid, usually in the form of DNA, is administered to modify the genetic repertoire of target cells for therapeutic purposes. This can be accomplished efficiently using a recombinant adenoviral vector encoding a so-called "therapeutic gene". A therapeutic gene is generally considered a gene that corrects or compensates for an underlying protein deficit or, alternately, a gene that is capable of down-regulating a particular gene, or counteracting the negative effects of its encoded product, in a given disease state or syndrome. Recombinant adenoviral vectors have been used to transfer one or more recombinant genes to diseased cells or tissues in need of treatment. As reviewed by Crystal, Science, 270, 404-410 (1995), such vectors are preferred over other vectors commonly employed for gene therapy (e.g., retroviral vectors) since adenoviral vectors can be produced in high titers (i.e., up to 10^{13} viral particles/ml), and they efficiently transfer genes to nonreplicating, as well as replicating, cells. Moreover, adenoviral vectors are additionally preferred based on their normal tropism for

the respiratory epithelium in cases where the targeted tissue for somatic gene therapy is the lung, as well as for other reasons (see, e.g., Straus, <u>In Adenoviruses</u>, Plenan Press, New York, NY, 451-496 (1984)); Horwitz et al., <u>In Virology</u>, 2nd Ed., Fields et al., eds., Raven Press, New York, NY, 1679-1721 (1990); Berkner, <u>BioTechniques</u>, <u>6</u>, 616 (1988); Chanock et al., <u>JAMA</u>, <u>195</u>, 151 (1966); Haj-Ahmad et al., <u>J. Virol.</u>, <u>57</u>, 267 (1986); and Ballay et al., <u>EMBO</u>, <u>4</u>, 3861 (1985)).

There are 49 human adenoviral serotypes, categorized into 6 subgenera (A through F) based on nucleic acid comparisons, fiber protein characteristics, and biological properties (Crawford-Miksza et al., <u>J. Virol.</u>, <u>70</u>, 1836-1844 (1996)). The group C viruses (e.g., serotypes 2 and 5, or Ad2 and Ad5) are well characterized. It is these serotypes that currently are employed for gene transfer studies, including human gene therapy trials (see, e.g., Rosenfeld et al., Science, 252, 431-434 (1991); Rosenfeld et al., <u>Cell</u>, <u>68</u>, 143-155 (1992); Zabner, <u>Cell</u>, <u>75</u>, 207-216 (1993); Crystal et al., Nat. Gen., 8, 42-51 (1994); Yei et al., Gene Therapy, 1, 192-200 (1994); Chen et al., Proc. Natl. Acad. Sci., 91, 3054-3057 (1994); Yang et al., Nat. Gen., 7, 362-369 (1994); Zabner et al., Nat. Gen., 6, 75-83 (1994)). Other groups and serotypes include, but are not limited to: group A (e.g., serotypes 12 and 31), group B (e.g., serotypes 3 and 7), group D (e.g., serotypes 8 and 30), group E (e.g., serotype 4) and group F (e.g., serotypes 40 and 41) (Horwitz et al., supra).

In terms of general structure, all adenoviruses examined to date are nonenveloped, regular icosahedrons of about 65 to 80 nanometers in diameter. Adenoviruses are comprised of linear, double-stranded DNA that is complexed with core proteins and surrounded by the adenoviral capsid. The capsid is comprised of 252 capsomeres, of which 240 are hexons and 12 are pentons. The hexon

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capsomere provides structure and form to the capsid (Pettersson, in <u>The Adenoviruses</u>, pp. 205-270, Ginsberg, ed., (Plenum Press, New York, NY, 1984)), and is a homotrimer of the hexon protein (Roberts et al., <u>Science</u>, 232, 1148-1151 (1986)). The penton comprises a penton base, which is bound to other hexon capsomeres, and a fiber, which is noncovalently bound to, and projects from, the penton base. The penton fiber protein comprises three identical polypeptides (i.e., polypeptide IV). The Ad2 penton base protein comprises five identical polypeptides (i.e., polypeptides, polypeptides, polypeptide).

The adenoviruses provide an elegant and efficient means of transferring therapeutic genes into cells. However, one problem encountered with the use of adenoviral vectors for gene transfer in vivo is the generation of antibodies to antigenic epitopes on adenoviral capsid proteins. If sufficient in titer, the antibodies can limit the ability of the vector to be used more than once as an effective gene transfer vehicle. For instance, animal studies demonstrate that intravenous or local administration (e.g., to the lung, heart or peritoneum) of an adenoviral type 2 or 5 gene transfer vector can result in the production of antibodies directed against the vector which prevent expression from the same serotype vector administered 1 to 2 weeks later (see, e.g., Yei et al., supra; Zabner (1994), supra; Setoguchi et al., Am. J. Respir. Cell. Mol. Biol., 10, 369-377 (1994); Kass-Eisler et al., Gene Therapy, 1, 395-402 (1994); Kass-Eisler et al., Gene Therapy 3, 154-162 (1996)). This is a drawback in adenoviral-mediated gene therapy, since many uses of an adenoviral vector (e.g., for prolonged gene therapy) require repeat administration inasmuch as the vector does not stably integrate into the host cell genome. The mechanism by which antibodies

directed against an adenovirus are able to prevent or reduce expression of an adenoviral-encoded gene is unclear. However, the phenomenon is loosely referred to as "neutralization", and the responsible antibodies are termed "neutralizing antibodies."

There are three capsid structures against which neutralizing antibodies potentially can be elicited: fiber, penton, and hexon (Pettersson, supra). The hexon protein, and to a lesser extent the fiber protein, comprise the main antigenic determinants of the virus, and also determine the serotype specificity of the virus (Watson et al., J. Gen. Virol., 69, 525-535 (1988); Wolfort et al., J. Virol., 62, 2321-2328 (1988); Wolfort et al., J. Virol., 56, 896-903 (1985); Crawford-Miksza et al., supra). Researchers have examined and compared the structure of these coat proteins of different adenoviral serotypes in an effort to define the regions of the proteins against which neutralizing antibodies are elicited.

The Ad2 hexon trimer is comprised of a pseudohexagonal base and a triangular top formed of three towers (Roberts et al., supra; Athappilly et al., J. Mol. Biol., 242, 430-455 (1994)). The base pedestal consists of two tightly packed eight-stranded antiparallel beta barrels stabilized by an internal loop. The predominant regions in hexon protein against which neutralizing antibodies are directed appear to be in loops 1 and 2 (i.e., LI or 11, and LII or 12, respectively) in one of the three towers. For instance, Kinloch et al. (J. Biol. Chem., 258, 6431-6436 (1984)) compared adenoviral hexon sequences and theorized that the serotype-specific antigenic determinants on hexon are located in amino acid residues 120 to 470 encompassing the 11 and 12 loops since type-specific sequence differences are mainly concentrated in this region. Toogood et al. (J. Gen.

vector (such as an adenovirus) comprising the chimeric protein to be administered and effect gene expression in the case where there are preexisting neutralizing antibodies directed against the wild-type adenovirus coat protein. The present invention also provides a vector, particularly an adenoviral vector, that comprises a chimeric adenovirus coat protein such as chimeric adenovirus hexon protein (and which optionally further comprises a chimeric adenovirus fiber and/or penton base protein), and methods of constructing and using such a vector.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 is a diagram of the method employed to construct the vector pAd70-100dlE3.fiber7.

Figure 2 is a partial restriction map of the vector pGBS.59-100(HSF:RGD).

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides, among other things, a chimeric adenovirus coat protein. The chimeric adenovirus coat protein comprises a nonnative amino acid sequence, such that the chimeric adenovirus coat protein (or a vector comprising the chimeric adenovirus coat protein) has a decreased ability or inability to be recognized by antibodies (e.g., neutralizing antibodies) directed against the corresponding wild-type adenovirus coat protein.

Chimeric Adenovirus Coat Protein

A "coat protein" according to the invention is either an adenoviral penton base protein, an adenoviral hexon protein, or an adenoviral fiber protein. Preferably a coat protein is a adenoviral hexon protein or an adenoviral fiber protein. Any one of the serotypes of

human or nonhuman adenovirus can be used as the source of the coat protein, or its gene or coding sequence.

Optimally, however, the adenovirus coat protein is that of a Group B or C adenovirus and, preferably, is that of Adl, Ad2, Ad3, Ad5, Ad6, Ad7, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, or Ad48.

The chimeric adenovirus coat protein (or a vector, such as adenoviral vector, comprising the chimeric adenovirus coat protein) has a decreased ability or an inability to be recognized by an antibody (e.g., a neutralizing antibody) directed against the corresponding wild-type adenovirus coat protein. A "neutralizing antibody" is an antibody that either is purified from or is present in serum. As used herein, an antibody can be a single antibody or a plurality of antibodies. An antibody is "neutralizing" if it inhibits infectivity of (i.e., cell entry) or gene expression commanded by an adenovirus comprising wild-type coat protein, or if it exerts a substantial deleterious effect on infectivity of or gene expression commanded by an adenovirus comprising wild-type coat protein, as compared, for instance, to any effect on any other adenoviral property.

An ability or inability of a chimeric coat protein to "be recognized by" (i.e., interact with) a neutralizing antibody directed against the wild-type adenovirus coat protein can be assessed by a variety of means known to those skilled in the art. For instance, the removal of one or more epitopes for a neutralizing antibody present in a wild-type adenovirus coat protein to generate a chimeric adenovirus coat protein will result in a decreased ability or inability of the chimeric coat protein to be recognized by the neutralizing antibody. Also, such a decreased ability or inability to interact with a neutralizing antibody directed against wild-type coat protein can be demonstrated by means of a

<u>Virol.</u>, <u>73</u>, 1429-1435 (1992)) used peptides from this region to generate specific anti-loop antisera and confirmed that antibodies against residues 281-292 of 11 and against residues 441-455 of 12 were sufficient to neutralize infection. Also, Crompton et al. (<u>J. Gen. Virol.</u>, <u>75</u>, 133-139 (1994)) modified these loops to accept neutralizing epitopes from polio virus, and demonstrated that infection with the resultant adenoviral vector generated neutralizing immunity against polio virus. More recently it was demonstrated that the hexon protein is composed of seven discrete hypervariable regions in loops and 1 and 2 (HVR1 to HVR7) which vary in length and sequence between adenoviral serotypes (Crawford-Miksza et al., <u>supra</u>).

Less is known regarding the regions of the fiber protein against which neutralizing antibodies potentially can be directed. However, much data is available on the structure of the fiber protein. The trimeric fiber protein consists of a tail, a shaft, and a knob (Devaux et al., J. Molec. Biol., 215, 567-588 (1990)). The fiber shaft region is comprised of repeating 15 amino acid motifs, which are believed to form two alternating beta strands and beta bends (Green et al., EMBO J., 2, 1357-1365 (1983)). The overall length of the fiber shaft region and the number of 15 amino acid repeats differ between adenoviral serotypes. The receptor binding domain of the fiber protein and sequences necessary for fiber trimerization are localized in the knob region encoded by roughly the last 200 amino acids of the protein (Henry et al., J. Virol., 68(8), 5239-5246 (1994)); Xia et al., Structure, 2(12), 1259-1270 (1994)). Furthermore, all adenovirus serotypes appear to possess a type of specific moiety located in the knob region (Toogood et al., supra.)

Given the existence of these potential epitopes in hexon protein and fiber protein, it is understandable

that, in some cases, difficulties have been encountered using adenovirus as a vector for gene therapy. Accordingly, recombinant adenoviral vectors capable of escaping such neutralizing antibodies (in the event they are preexisting and hamper gene expression commanded by adenovirus in an initial dose), and which would allow repeat doses of adenoviral vectors to be administered, would significantly advance current gene therapy methodology.

Thus, the present invention seeks to overcome at least some of the aforesaid problems of recombinant adenoviral gene therapy. In particular, it is an object of the present invention to provide a recombinant adenovirus comprising a chimeric coat protein that has a decreased ability or inability to be recognized by antibodies (i.e., neutralizing antibodies) directed against the corresponding wild-type adenovirus coat protein. These and other objects and advantages of the present invention, as well as additional inventive features, will be apparent from the description of the invention provided herein.

BRIEF SUMMARY OF THE INVENTION

The present invention provides a chimeric adenovirus coat protein (particularly a chimeric adenovirus hexon protein) comprising a nonnative amino acid sequence. The chimeric adenovirus coat protein is not recognized by, or has a decreased ability to be recognized by, a neutralizing antibody directed against the corresponding wild-type (i.e., native) coat protein. The chimeric adenovirus coat protein enables a vector (such as an adenovirus) comprising the corresponding protein to be administered repetitively, or to be administered following administration of an adenovirus vector comprising the corresponding wild-type coat protein. It also enables a

neutralization test (see, e.g., Toogood et al., <u>supra;</u> Crawford-Miksza et al., <u>supra;</u> Mastrangeli et al., <u>Human</u> <u>Gene Therapy</u>, <u>7</u>, 79-87 (1996)), or as further described herein.

Generally, an "inability" of a chimeric adenovirus coat protein (or a vector comprising a chimeric adenovirus coat protein) to be recognized by a neutralizing antibody directed against wild-type adenovirus coat protein means that such an antibody does not interact with the chimeric coat protein, and/or exhibits no substantial deleterious effect on infectivity of or gene expression commanded by an adenovirus comprising wild-type coat protein, as compared, for instance, to any effect on any other adenoviral property.

A "decreased ability" to be recognized by neutralizing antibody directed against wild-type adenovirus coat protein refers to any decrease in the ability of the chimeric adenovirus coat protein (or a vector comprising the chimeric coat protein) to be recognized by an antibody directed against the corresponding wild-type adenovirus coat protein as compared to the wild-type adenovirus coat protein. such ability/inability is assessed by means of a neutralization test in particular, preferably a "decreased ability" to be recognized by a neutralizing antibody directed against wild-type adenovirus coat protein is exhibited by from about a 10% to about a 99% increase in the ability of a recombinant adenovirus comprising the chimeric coat protein to cause a visible cytopathic effect (c.p.e.) in cells such as A549 cells or COS-1 cells (or other such cells appropriate for a neutralization assay) in the presence of the neutralizing antibody as compared to an adenovirus comprising the wild-type coat protein against which the neutralizing antibody is directed.

Furthermore, a decreased ability or inability of an adenovirus chimeric coat protein (or a vector comprising a chimeric adenovirus coat protein) to interact with a neutralizing antibody can be shown by a reduction of inhibition (from about 10% to about 99%) or no inhibition at all of cell infectivity by a recombinant vector (such as an adenoviral vector) containing the chimeric coat protein as compared to a recombinant vector containing the wild-type protein. Also, a decreased ability or inability of an adenovirus chimeric coat protein (or a vector comprising a chimeric adenovirus coat protein) to interact with a neutralizing antibody can be shown by a reduction of inhibition (from about 10% to about 99%) or no inhibition at all of gene expression commanded by a recombinant vector (such as an adenoviral vector) containing the chimeric coat protein as compared to a recombinant vector containing the wild-type coat protein. These tests can be carried out when the recombinant adenovirus containing the chimeric coat protein is administered following the administration of an adenovirus containing the wild-type coat protein, or when the recombinant adenovirus is administered to a host that has never before encountered or internalized adenovirus (i.e., a "naïve" host). These methods are described, for instance, in the Examples which follow as well as in Mastrangeli et al., supra. Other means such as are known to those skilled in the art also can be employed.

The coat protein is "chimeric" in that it comprises a sequence of amino acid residues that is not typically found in the protein as isolated from, or identified in, wild-type adenovirus, which comprises the so-called native coat protein, or "wild-type coat protein". The chimeric coat protein thus comprises (or has) a "nonnative amino acid sequence" is meant any amino acid sequence (i.e., either component

residues or order thereof) that is not found in the native coat protein of a given serotype of adenovirus, and which preferably is introduced into the coat protein at the level of gene expression (i.e., by production of a nucleic acid sequence that encodes the nonnative amino acid sequence). Generally, the nonnative amino acid sequence can be obtained by deleting a portion of the amino acid sequence, deleting a portion of the amino acid sequence and replacing the deleted amino sequence with a so-called "spacer region", or introducing the spacer region into an unmodified coat protein. Preferably such manipulations result in a chimeric adenovirus coat protein according to the invention that is capable of carrying out the functions of the corresponding wild-type adenovirus coat protein (or, at least that when incorporated into an adenovirus, will allow appropriate virion formation and will not preclude adenoviral-mediated cell entry), and, optimally, that is not impeded in its proper folding. Also, it is desirable that the manipulations do not result in the creation of new epitopes for differing antibodies, unless, of course, such epitopes do not interfere with use of an adenovirus containing the chimeric coat protein as a gene transfer vehicle in vivo.

In particular, a nonnative amino acid sequence according to the invention preferably comprises a deletion of a region of a wild-type adenovirus coat protein, particularly an adenovirus hexon or fiber protein.

Optimally the resultant nonnative amino acid sequence is such that one or more of the existing epitopes for neutralizing antibodies directed against the corresponding wild-type adenovirus coat protein have been rendered non-immunogenic. Desirably, the region deleted comprises from about 1 to about 750 amino acids, preferably from about 1 to about 500 amino acids, and optimally from about 1 to about 300 amino acids. It also is desirable that the

region deleted comprises a smaller region less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids. The chimeric coat protein also desirably comprises a plurality of such deletions. Thus, according to the invention, the chimeric adenovirus coat protein comprises modification of one or more amino acids, and such modification is made in one or more regions.

In a preferred embodiment of the present invention, a nonnative amino acid sequence comprises a deletion of one or more regions of a wild-type adenovirus hexon protein, wherein preferably the hexon protein is the Ad2 hexon protein [SEQ ID NO:2] (which is encoded by the sequence of SEQ ID NO:1; GenBank® Data Bank Accession Number U20821), or the Ad5 hexon protein [SEQ ID NO:3] (GenBank® Data Bank Accession Number M73260, which is encoded by the sequence of SEQ ID NO:4), or the Ad7 hexon protein (GenBank® Data Bank Accession Number x76551). Alternately, preferably the hexon protein is the protein sequence reported by Crawford-Miksza et al. (Ad2 hexon [SEQ ID NO:52], Ad5 hexon SEQ ID NO:54]). In particular, the sequences of Crawford-Miksza et al. differ over those reported in the GenBank® Data Bank in that the amino acid residue reported as the first in the Crawford-Miksza et al. sequences is not Met, and the Ad5 hexon sequence is reported as terminating with "Gln His" instead of with "Thr Thr". As employed herein, the numbering of adenovirus hexon amino acid residues corresponds to that in Crawford-Miksza et al.

Desirably the region(s) of the deletion comprises an internal hexon protein sequence ("internal" meaning not at or near the C- or N-terminus of the protein; "near" referring to a distance of 500 amino acids or less), preferably a hypervariable region, e.g., as reported in Crawford-Miksza et al. In particular, optimally, the

internal region of the wild-type hexon protein that is deleted to generate the chimeric hexon protein comprises the entirety of 11 loop, preferably from about residue 131 to about residue 331 of the Ad2 hexon protein [SEQ ID NO:6] (which is encoded by the sequence of SEQ ID NO:5), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad5 [SEQ ID NO:8] (which is encoded by the sequence of SEQ ID NO:7), Ad6, Ad7, Ad8, Ad12, Ad16, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

Alternately, preferably the internal region of the wild-type hexon protein that is deleted to produce the chimeric hexon protein comprises one or more regions (e.g., smaller regions) of the 11 loop. Optimally the region deleted comprises a hypervariable region. Desirably the one or more regions of the 11 loop deleted are regions (i.e., hypervariable regions) selected from this group consisting of the HVR1 region, the HVR2 region, the HVR3 region, the HVR4 region, the HVR5 region, and the HVR6 region. Moreover, preferably the region of the wildtype protein that is deleted (or otherwise manipulated as described herein) occurs on the external surface of the hexon protein. Thus, HVR2, HVR3, HVR4, and HVR5 -- each of which are externally located regions of the hexon protein -- are particularly preferred for deletion or modification.

The "HVR1 region" preferably comprises from about amino acid 137 to about amino acid 188 of the Ad2 hexon protein [SEQ ID NO:10] (which is encoded by the sequence of SEQ ID NO:9), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:12] (which is encoded by the sequence of SEQ ID NO:11), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48,

BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR2 region" preferably comprises from about amino acid 194 to about amino acid 204 of the Ad2 hexon protein [SEQ ID NO:14] (which is encoded by the sequence of SEQ ID NO:13), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:16] (which is encoded by the sequence of SEQ ID NO:15), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR3 region" preferably comprises from about amino acid 222 to about amino acid 229 of the Ad2 hexon protein [SEQ ID NO:18] (which is encoded by the sequence of SEQ ID NO:17), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:20] (which is encoded by the sequence of SEQ ID NO:19), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR4 region" preferably comprises from about amino acid 258 to about amino acid 271 of the Ad2 hexon protein [SEQ ID NO:22] (which is encoded by the sequence of SEQ ID NO:21), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:24] (which is encoded by the sequence of SEQ ID NO:23), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR5 region" preferably comprises from about amino acid 278 to about amino acid 294 of the Ad2 hexon protein [SEQ ID NO:26] (which is encoded by the sequence

of SEQ ID NO:25), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Adl, Ad3, Ad5 [SEQ ID NO:28] (which is encoded by the sequence of SEQ ID NO:27), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra. In particular, preferably the deleted region comprises from about amino acid 297 to about amino acid 304 just outside of the HVR5 region of the Ad2 hexon protein [SEQ ID NO:30] (which is encoded by the sequence of SEQ ID NO:29), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Adl, Ad3, Ad5 [SEQ ID NO:32] (which is encoded by the sequence of SEQ ID NO:31), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR6 region" preferably comprises from about amino acid 316 to about amino acid 327 of the Ad2 hexon protein [SEQ ID NO:34] (which is encoded by the sequence of SEQ ID NO:33), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:36] (which is encoded by the sequence of SEQ ID NO:35), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

In another preferred embodiment of the invention, the internal region of the wild-type hexon protein that is deleted to generate the chimeric hexon protein comprises the entirety of the 12 loop, preferably from about residue 423 to about residue 477 of the Ad2 hexon protein [SEQ ID NO:38] (which is encoded by the sequence of SEQ ID NO:37), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from

Ad1, Ad3, Ad5 [SEQ ID NO:40] (which is encoded by the sequence of SEQ ID NO:39), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra. Alternately, preferably the internal region of the wild-type hexon protein that is deleted to produce the chimeric hexon protein comprises one or more smaller regions (e.g., hypervariable regions) of the 12 loop. In particular, preferably the smaller region of the 12 loop comprises the HVR7 region.

The "HVR7 region" preferably comprises from about amino acid 433 to about amino acid 465 of the Ad2 hexon protein [SEQ ID NO:42] (which is encoded by the sequence of SEQ ID NO:41), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Adl, Ad3, Ad5 [SEQ ID NO:44] (which is encoded by the sequence of SEQ ID NO:43), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra. In particular, preferably the deleted region comprises from about amino acid 460 to about amino acid 466 of the HVR7 region (i.e., extending one base pair outside of this region) of the Ad2 hexon protein [SEQ ID NO:46] (which is encoded by the sequence of SEQ ID NO:45), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:48] (which is encoded by the sequence of SEQ ID NO:47), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

Along the same lines, the chimeric adenovirus hexon protein desirably comprises deletions in one or both of the aforementioned regions, i.e., the hexon protein comprises deletions in one or both of the 11 and 12 loops,

which deletions can constitute the entirety of the loop(s), or can comprise deletions of one or more smaller regions (e.g., hypervariable regions) in one or both of the hexon loops. In particular, desirably the deleted region(s) are selected from the group consisting of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48, and equivalents and conservative variations of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48.

An "equivalent" is a naturally occurring variation of an amino acid or nucleic acid sequence, e.g., as are observed among different strains of adenovirus. A conservative variation is a variation of an amino acid sequence that results in one or more conservative amino acid substitution(s). A "conservative amino acid substitution" is an amino acid substituted by an alternative amino acid of similar charge density, hydrophilicity/hydrophobicity, size, and/or configuration (e.g., basic, Arg and Lys; aliphatic Ala, Cys, Gly, Ile, Leu, Met and Val; aromatic, Phe, Tyr, Trp, and His; hydrophilic, Glu, Gln, Asn, and Asp; hydroxyl, Ser and Thr).

In another preferred embodiment, the nonnative amino acid sequence of the chimeric adenoviral coat protein (i.e., particularly a chimeric adenoviral fiber or hexon protein) comprises a deletion of one or more region(s) of the wild-type adenovirus coat protein (particularly the 11

and/or 12 loops, and, most particularly, the HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, and/or HVR7 regions of the wildtype adenovirus hexon protein) as previously described, and further comprises a replacement of the region(s) with a spacer region preferably of from 1 to about 750 amino acids, especially of from about 1 to about 500 amino acids, and particularly of from about 1 to about 300 amino It also is desirable that the region deleted and replaced comprises a smaller region less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids. The chimeric coat protein also desirably comprises a plurality of such replacements. Thus, according to the invention, the chimeric adenovirus coat protein comprises modification of one or more amino acids, and such modification is made in one or more regions which can be a smaller region. A spacer region of the aforementioned size also preferably simply can be inserted into one of the aforementioned regions (particularly into the 11 and/or 12 loop, or one or more of the aforementioned HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, and HVR7 regions of the adenovirus hexon protein) in the absence of any deletion to render the resultant chimeric protein nonimmunogenic by, for instance, destroying the ability of a neutralizing antibody to interact with that particular site (e.g., by changing the spatial juxtaposition of critical amino acids with which the antibody interacts).

Optimally the spacer region comprises a nonconservative variation of the amino acid sequence of wild-type adenovirus coat protein (particularly wild-type adenovirus hexon protein) that comprises an epitope for a neutralizing antibody, and which may or may not be deleted upon the insertion of the spacer region. A "nonconservative variation" is a variation of this amino acid sequence that does not result in the creation or

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recreation in the chimeric adenovirus coat protein of the epitope for a neutralizing antibody directed against the wild-type adenovirus coat protein, and, in particular, is a variation of the spacer region that results in one or more nonconservative amino acid insertion(s) or substitution(s) in this region. A "nonconservative amino acid substituted by an alternative amino acid of differing charge density, hydrophilicity/hydrophobicity, size, and/or configuration (e.g., a change of a basic amino acid for an acidic amino acid, a hydrophilic amino acid for a hydrophobic amino acid, and the like).

Desirably the spacer region does not interfere with the functionality of the chimeric adenovirus coat protein, particularly the chimeric adenovirus hexon or fiber protein, e.g., the ability of hexon protein to bind penton base protein or other hexon capsomeres, or the ability of penton fiber to bind penton base and/or to a cell surface receptor. Such functionality can be assessed by virus viability. Similarly, the absence of the creation or recreation of the epitope(s) for a neutralizing antibody directed against the wild-type coat (e.g., hexon and/or fiber) protein can be confirmed using techniques as described in the Examples which follow (e.g., by ensuring the antibody, which may be in a carrier fluid such as serum or other liquid, binds the wild-type adenovirus coat protein, but not the chimeric adenovirus coat protein).

Preferably the spacer region incorporated into the adenovirus coat protein (i.e., either as an insertion into the wild-type coat protein, or to replace one or more deleted region(s) of the wild-type adenovirus coat protein) comprise a series of polar and/or charged amino acids (e.g., Lys, Arg, His, Glu, Asp, and the like), or amino acids with intermediate polarity (e.g., Gln, Asn, Thr, Ser, Met, and the like). In particular, desirably

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the spacer region comprises the sequence of SEQ ID NO:50 (which is encoded by the sequence of SEQ ID NO:49), and equivalents and conservative variations of SEQ ID NO:50. Alternately, the spacer region can comprise any other sequence like the FLAG octapeptide sequence of SEQ ID NO:50 that will not interfere with the functionality of the resultant chimeric protein.

In still yet another preferred embodiment, a region of a wild-type adenovirus coat protein (particularly an adenovirus hexon and/or fiber protein) is deleted and replaced with a spacer region comprising the corresponding coat protein region of another adenoviral serotype. Preferably in this embodiment the spacer region is of a different adenoviral group. For instance, preferably a region of an Ad2 coat protein can be replaced with the corresponding region of an Ad5 or Ad7 coat protein (or any other serotype of adenovirus as described above), and vice versa. It also is preferable that such a spacer region comprising the coat protein region of another adenoviral serotype is simply inserted into the corresponding coat protein region of the chimeric coat protein. In this case, the likelihood of obtaining a chimeric hexon protein that is functional can be increased by making sure that the size of the hypervariable domain resulting from such insertion approximates the size of a known hypervariable domain. For instance, the HVR1 region of Ad40 is about 30 amino acids smaller than the HVR1 region of Ad2 (as well as other adenoviruses such as Ad5, Ad8, etc.). Thus, preferably a spacer region of about 30 amino acids can be incorporated into the Ad40 HVR1 region to produce a chimeric adenovirus hexon protein. In particular, desirably the region of Ad2 (or other adenovirus) that is not present in Ad40 (i.e., approximately amino acid residues 138 to 174), or a portion thereof, is introduced

into Ad40 to produce the chimeric adenoviral hexon protein.

According to the invention, desirably the nonnative amino acid sequence of a chimeric coat protein comprises a plurality of such replacements or insertions. When the coat protein is incorporated into an adenoviral vector, preferably the entire coat protein of one adenoviral serotype can be substituted with the entire coat protein of another adenoviral serotype, as described further herein.

The region or regions of wild-type adenovirus hexon protein that are deleted and replaced by the spacer region, or into which the spacer region is inserted, can be any suitable region(s) and desirably comprise one or more of the regions described above with respect to the hexon protein deletions. For instance, preferably the one or more regions into which the spacer region is inserted or which the spacer region replaces comprises the entirety of the 11 and/or 12 loop, or a sequence selected from the group consisting of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48, and equivalents and conservative variations of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48.

Similarly, the spacer region itself (i.e., both for insertion as well as replacement) preferably comprises the entirety of the 11 and/or 12 loop, or a sequence selected

from the group consisting of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48, and equivalents and conservative variations of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:36, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48.

The fiber protein also preferably is altered in a similar fashion as described for modification of hexon protein to escape antibodies directed in particular against wild-type adenovirus fiber protein. protein sequences and methods of modifying fiber protein are known to those skilled in the art (see, e.g., Xia et al., supra; Novelli et al., Virology, 185, 365-376 (1991)). The fiber manipulations can be carried out in the absence of, or along with, modifications to the adenovirus hexon protein. In particular, preferably the fiber protein can be replaced in its entirety, or in part, with sequences of a fiber protein from a different serotype of adenovirus. Also, preferably, deletions can be made of fiber sites that constitute an epitope for a neutralizing antibody, and/or insertions can be made at the site to destroy the ability of the protein to interact with the antibody.

Nucleic Acid Encoding The Chimeric Adenovirus Coat Protein

Preferably the chimeric adenovirus coat protein (particularly the chimeric adenovirus hexon or fiber protein) comprises a nonnative amino acid sequence wherein

the alteration is made at the level of DNA. Thus, the invention preferably provides an isolated and purified nucleic acid encoding a chimeric adenovirus coat protein. Desirably, the invention provides an isolated and purified nucleic acid encoding a chimeric adenovirus hexon protein as defined herein, wherein the nucleic acid sequence comprises a deletion of a region (or a plurality of such deletions) that encodes from about 1 to about 750 amino acids of the wild-type adenovirus coat protein, preferably from about 1 to about 500 amino acids, and optimally from about 1 to about 300 amino acids. It also is desirable that the region deleted comprises a smaller region that encodes less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids. In particular, optimally the deletion (e.g., of an adenoviral hexon protein) comprises the entirety of the 11 and/or 12 loop, or a sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, and SEQ ID NO:47, or a sequence comprising the corresponding region from Adl, Ad3, Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The invention also preferably provides an isolated and purified nucleic acid encoding a chimeric adenovirus hexon protein as defined herein, wherein the nucleic acid sequence comprises a deletion of one or more sequences selected from the group consisting of equivalents and conservatively modified variants of sequences that encode the entirety of the 11 and/or 12 loop, or SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID

NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, and SEQ ID NO:47, or a sequence comprising the corresponding region from Ad1, Ad3, Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

With respect to the nucleic acid sequence, an "equivalent" is a variation on the nucleic acid sequence such as can occur in different strains of adenovirus, and which either does or does not result in a variation at the amino acid level. Failure to result in variation at the amino acid level can be due, for instance, to degeneracy in the triplet code. A "conservatively modified variant" is a variation on the nucleic acid sequence that results in one or more conservative amino acid substitutions. In comparison, a "nonconservatively modified variant" is a variation on the nucleic acid sequence that results in one or more nonconservative amino acid substitutions.

In another preferred embodiment, the invention provides an isolated and purified nucleic acid encoding a chimeric adenovirus coat protein wherein the nucleic acid sequence further comprises a replacement of the deleted region (or a plurality of such replacements) with a spacer nucleic acid region (i.e., the nucleic acid sequence that encodes the aforementioned "spacer region") that encodes from about 1 to about 750 amino acids of the wild-type adenovirus coat protein, preferably from about 1 to about 300 amino acids, and optimally from about 1 to about 300 amino acids. It also is desirable that the region deleted and replaced comprises a smaller region that encodes less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids.

Preferably, the spacer nucleic acid region comprises a FLAG octapeptide-encoding sequence [SEQ ID NO:49], and equivalents and conservatively modified variants of SEQ ID NO:49. Similarly, a spacer nucleic acid region can be employed that substitutes one or more coat protein encoding regions (particularly a hexon protein encoding region) of a particular adenoviral serotype with a coat protein encoding region (particularly a hexon protein encoding region) of another adenoviral serotype. Thus, preferably a spacer nucleic acid region present in a chimeric adenoviral hexon protein is selected from the group consisting of sequences that encode the entirety of the 11 and/or 12 loop, or SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, and SEQ ID NO:47, or a sequence comprising the corresponding region from Adl, Ad3, Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra, and equivalents and conservatively modified variants of these sequences.

As described above with respect to the chimeric adenovirus coat protein, the spacer nucleic acid region (or a plurality thereof) simply can be incorporated into the coat protein in the absence of any deletions. These manipulations can be carried out so as to produce the above-described chimeric adenovirus coat protein.

The means of making such a chimeric adenoviral coat protein (i.e., by introducing conservative or nonconservative variations at either the level of DNA or protein) are known in the art, are described in the Examples which follow, and also can be accomplished by means of various commercially available kits and vectors

(e.g., New England Biolabs, Inc., Beverly, MA; Clontech, Palo Alto, CA; Stratagene, LaJolla, CA, and the like). In particular, the ExSite™ PCR-based site-directed mutagenesis kit and the Chameleon™ double-stranded site-directed mutagenesis kit by Stratagene can be employed for introducing such mutations. Moreover, the means of assessing such mutations (e.g., in terms of effect on ability not to be neutralized by antibodies directed against wild-type hexon protein) are described in the Examples herein.

Accordingly, the present invention provides a preferred means of making a chimeric adenoviral coat protein, particularly a chimeric adenoviral hexon protein, which comprises obtaining an adenoviral genome encoding the wild-type adenovirus coat protein (e.g., the wild-type adenovirus hexon protein), and deleting one or more region(s) of the chimeric adenovirus coat protein (particularly the chimeric adenovirus hexon protein) comprising from about 1 to about 750 amino acids by modifying the corresponding nucleic acid coding sequence. Similarly, the invention provides a method of making a chimeric adenovirus coat protein (particularly a chimeric adenovirus hexon protein) which comprises obtaining an adenoviral genome encoding the wild-type adenovirus coat protein, deleting one or more region(s) of the adenovirus coat protein comprising from about 1 to about 750 amino acids by modifying the corresponding coding sequence, and replacing the deleted region(s) with a spacer region comprising from about 1 to about 300 amino acids by introducing a nucleic acid region (i.e., a "spacer nucleic acid region") that codes for same. Alternately, the spacer region preferably is simply incorporated into the coat protein (particularly the hexon protein) in the absence of any deletion. Optimally the spacer nucleic acid region encodes a nonconservative variation of the

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amino acid sequence of the wild-type adenovirus coat protein. The size of the DNA used to replace the native coat protein coding sequence may be constrained, for example, by impeded folding of the coat protein or improper assembly of the coat protein into a complex (e.g., penton base/hexon complex) or virion. DNA encoding 150 amino acids or less is particularly preferred for insertion/replacement in the chimeric coat protein gene sequence, and DNA encoding 50 amino acids or less is even more preferred.

Briefly, the method of mutagenesis comprises deleting one or more regions of an adenovirus coat protein, and/or inserting into an adenovirus coat protein one or more regions with a differing amino acid sequence, particularly by manipulating the DNA sequence. Several methods are available for carrying out such manipulations of adenovirus coat protein DNA sequences; these methods further can be used in combination. The method of choice depends on factors known to those skilled in the art, e.g., the size of the DNA region to be manipulated. For instance, convenient restriction sites (which further can be introduced into a sequence) can be used to introduce or remove segments of DNA, or entire genes or coding sequences. Alternately, other methods of mutagenesis involve the hybridization of a mismatched oligonucleotide to a region of single-stranded target DNA, extending the primer, for instance, using T7 DNA polymerase or other such means to produce a double-stranded heteroduplex, and isolating the mutant strand that incorporates the mismatched oligonucleotide from the parental nonmutant strand for use as a template and in further manipulations. The mutant strand can be separated from the parental strand using various selection means known to those skilled in the art (see, e.g., Kunkel et al., Methods Enzymol., 204, 125-139 (1991), as well as the underlying

methodology employed in the Chameleon™ kit). Alternately, the parental strand can be selectively degraded, for instance, with use of enzymes that nick the nonmethylated strand of a hemi-methylated DNA molecule (e.g., HpaII, MspI, and Sau3AI), and by extending the mutant strand using 5-methyl-dCTP, which renders the strand resistant to cleavage by these enzymes. Along the same lines, an entirely PCR-based approach can be employed for making mutations (e.g., Kunkel, Proc. Natl. Acad. Sci., 82, 488-492 (1985); Costa et al., Nucleic Acids Res., 22, 2423 (1994)), for instance, such as the approach encompassed by the $\operatorname{ExSite}^{\operatorname{TM}}$ kit. More generally, amino acid substitutions or deletions can be introduced during PCR by incorporating appropriate mismatches in one or both primers. Once the chimeric coat protein sequence has been produced, the nucleic acid fragment encoding the sequence further can be isolated, e.g., by PCR amplification using 5' and 3' primers, or through use of convenient restriction sites.

Vector Comprising a Chimeric Hexon Protein

A "vector" according to the invention is a vehicle for gene transfer as that term is understood by those skilled in the art, and includes viruses, plasmids, and the like. A preferred vector is an adenovirus, particularly a virus of the family Adenoviridae, and desirably of the genus Mastadenovirus (e.g., comprised of mammalian adenoviruses) or Aviadenovirus (e.g., comprised of avian adenoviruses). Such an adenovirus (or other viral vector) can be transferred by its own means of effecting cell entry (e.g., by receptor-mediated endocytosis), or can be transferred to a cell like a plasmid, i.e., in the form of its nucleic acid, for instance, by using liposomes to transfer the nucleic acid, or by microinjecting or transforming the DNA into the cell. The nucleic acid vectors that can be employed for

gene transfer, particularly the adenoviral nucleic acid vectors, are referred to herein as "transfer vectors". Such nucleic acid vectors also include intermediary plasmid vectors that are employed, e.g., in the construction of adenoviral vectors.

Desirably an adenoviral vector is a serotype group C virus, preferably an Ad2 or Ad5 vector, although any other serotype adenoviral vector (e.g., group A including serotypes 12 and 31, group B including serotypes 3 and 7, group D including serotypes 8 and 30, group E including serotype 4, and group F including serotypes 40 and 41, and other Ad vectors previously described) can be employed. An adenoviral vector employed for gene transfer can be replication competent. Alternately, an adenoviral vector can comprise genetic material with at least one modification therein, which renders the virus replication deficient. The modification to the adenoviral genome can include, but is not limited to, addition of a DNA segment, rearrangement of a DNA segment, deletion of a DNA segment, replacement of a DNA segment, or introduction of a DNA lesion. A DNA segment can be as small as one nucleotide and as large as 36 kilobase pairs (i.e., the approximate size of the adenoviral genome) or, alternately, can equal the maximum amount which can be packaged into an adenoviral virion (i.e., about 38 kb). Preferred modifications to the group C adenoviral genome include modifications in the E1, E2, E3 and/or E4 regions. Similarly, an adenoviral vector can be a cointegrate, i.e., a ligation of adenoviral sequences with other sequences, such as other virus sequences, particularly baculovirus sequences, or plasmid sequences, e.g., so as to comprise a prokaryotic or eukaryotic expression vector.

In terms of an adenoviral vector (particularly a replication deficient adenoviral vector), such a vector can comprise either complete capsids (i.e., including a

viral genome such as an adenoviral genome) or empty capsids (i.e., in which a viral genome is lacking, or is degraded, e.g., by physical or chemical means). The capsid further can comprise nucleic acid linked to the surface by means known in the art (e.g., Curiel et al., Human Gene Therapy, 3, 147-154 (1992)) or can transfer non-linked nucleic acid, for instance, by adenoviral-mediated uptake of bystander nucleic acid (e.g., PCT International Application WO 95/21259).

Along the same lines, since methods are available for transferring an adenovirus in the form of its nucleic acid sequence (i.e., DNA), a vector (i.e., a transfer vector) similarly can comprise DNA, in the absence of any associated protein such as capsid protein, and in the absence of any envelope lipid. Inasmuch as techniques are available for making a RNA copy of DNA (e.g., in vitro transcription), and inasmuch as RNA viruses also can be employed as vectors or transfer vectors, a transfer vector also can comprise RNA. Thus, according to the invention whereas a vector comprises (and, further, may encode) a chimeric adenoviral coat protein, a transfer vector typically encodes a chimeric adenoviral coat protein (particularly a chimeric adenoviral hexon and/or fiber protein).

Based on this, the invention provides an adenoviral vector that comprises a chimeric coat protein (particularly a chimeric hexon and/or fiber protein) according to the invention. Preferably such a vector comprises a chimeric coat protein (particularly a chimeric adenovirus hexon protein and/or chimeric adenovirus fiber protein) as described above. Alternately, preferably the vector lacks wild-type fiber protein, e.g., the vector encodes a truncated or non-functional fiber protein, or fails to translate fiber protein. Such fiber mutations and the means of introducing fiber mutations are known to

those skilled in the art (see, e.g., Falgout et al., \underline{J} . Virol., 62, 622-625 (1988)).

Of course, the chimeric adenoviral coat proteins include coat proteins in which the native (i.e., wild-type) hexon and/or fiber protein of an adenoviral vector is replaced by a hexon and or fiber amino acid sequence of a different adenoviral serotype such that the resultant adenoviral vector has a decreased ability or inability to be recognized by neutralizing antibodies directed against the corresponding wild-type coat protein. This replacement can comprise the entirety of the hexon and/or fiber amino acid sequence, or only a portion, as described above. Both proteins can be manipulated (e.g., in a single adenovirus), or only a single chimeric adenovirus coat protein can be employed, with the remaining coat proteins being wild-type.

A vector according to the invention (including a transfer vector) preferably comprises additional sequences and mutations, e.g., some that can occur within the coat protein coding sequence itself. In particular, a vector according to the invention further preferably comprises a nucleic acid encoding a passenger gene or passenger coding sequence. A "nucleic acid" is a polynucleotide (i.e., DNA or RNA). A "gene" is any nucleic acid sequence coding for a protein or an RNA molecule. Whereas a gene comprises coding sequences plus any non-coding sequences, a "coding sequence" does not include any non-coding (e.g., regulatory) DNA. A "passenger gene" or "passenger coding sequence" is any gene which is not typically present in and is subcloned into a vector (e.g., a transfer vector) according to the present invention, and which upon introduction into a host cell is accompanied by a discernible change in the intracellular environment (e.g., by an increased level of deoxyribonucleic acid (DNA), ribonucleic acid (RNA), peptide or protein, or by an

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altered rate of production or degradation thereof). A "gene product" is either an as yet untranslated RNA molecule transcribed from a given gene or coding sequence (e.g., mRNA or antisense RNA) or the polypeptide chain (i.e., protein or peptide) translated from the mRNA molecule transcribed from the given gene or coding sequence. A gene or coding sequence is "recombinant" if the sequence of bases along the molecule has been altered from the sequence in which the gene or coding sequence is typically found in nature, or if the sequence of bases is not typically found in nature. According to this invention, a gene or coding sequence can be naturally occurring or wholly or partially synthetically made, can comprise genomic or complementary DNA (cDNA) sequences, and can be provided in the form of either DNA or RNA.

Non-coding sequences or regulatory sequences include promoter sequences. A "promoter" is a DNA sequence that directs the binding of RNA polymerase and thereby promotes RNA synthesis. "Enhancers" are cis-acting elements of DNA that stimulate or inhibit transcription of adjacent genes. An enhancer that inhibits transcription is also termed a "silencer". Enhancers differ from DNA-binding sites for sequence-specific DNA binding proteins found only in the promoter (which also are termed "promoter elements") in that enhancers can function in either orientation, and over distances of up to several kilobase pairs, even from a position downstream of a transcribed region. According to the invention, a coding sequence is "operably linked" to a promoter (e.g., when both the coding sequence and the promoter constitute a passenger gene) when the promoter is capable of directing transcription of that coding sequence.

Accordingly, a "passenger gene" can be any gene, and desirably either is a therapeutic gene or a reporter gene. Preferably a passenger gene is capable of being expressed

in a cell in which the vector has been internalized. instance, the passenger gene can comprise a reporter gene, or a nucleic acid sequence which encodes a protein that can be detected in a cell in some fashion. The passenger gene also can comprise a therapeutic gene, for instance, a therapeutic gene which exerts its effect at the level of RNA or protein. Similarly, a protein encoded by a transferred therapeutic gene can be employed in the treatment of an inherited disease, such as, e.g., the cystic fibrosis transmembrane conductance regulator cDNA for the treatment of cystic fibrosis. The protein encoded by the therapeutic gene can exert its therapeutic effect by resulting in cell killing. For instance, expression of the gene in itself may lead to cell killing, as with expression of the diphtheria toxin A gene, or the expression of the gene may render cells selectively sensitive to the killing action of certain drugs, e.g., expression of the HSV thymidine kinase gene renders cells sensitive to antiviral compounds including acyclovir, gancyclovir and FIAU (1-(2-deoxy-2-fluoro-b-Darabinofuranosil)-5-iodouracil). Moreover, the therapeutic gene can exert its effect at the level of RNA, for instance, by encoding an antisense message or ribozyme, by affecting splicing or 3' processing (e.g., polyadenylation), or by encoding a protein which acts by affecting the level of expression of another gene within the cell (i.e., where gene expression is broadly considered to include all steps from initiation of transcription through production of a processed protein), perhaps, among other things, by mediating an altered rate of mRNA accumulation, an alteration of mRNA transport, and/or a change in post-transcriptional regulation. Accordingly, the use of the term "therapeutic gene" is intended to encompass these and any other embodiments of that which is more commonly referred to as gene therapy

and is known to those of skill in the art. Similarly, the recombinant adenovirus can be used for gene therapy or to study the effects of expression of the gene (e.g., a reporter gene) in a given cell or tissue in vitro or in vivo, or for diagnostic purposes.

Also, a passenger coding sequence can be employed in the vector. Such a coding sequence can be employed for a variety of purposes even though a functional gene product may not be translated from the vector sequence. For instance, the coding sequence can be used as a substrate for a recombination reaction, e.g., to recombine the sequence with the host cell genome or a vector resident in the cell. The coding sequence also can be an "anticoding sequence," e.g., as appropriate for antisense approaches. Other means of using the coding sequence will be known to one skilled in the art.

The present invention thus provides recombinant adenoviruses comprising a chimeric hexon protein and/or a chimeric fiber protein, and which preferably additionally comprise a passenger gene or genes capable of being expressed in a particular cell. The recombinant adenoviruses can be generated by use of a vector, specifically, a transfer vector, and preferably a viral (especially an adenoviral) or plasmid transfer vector, in accordance with the present invention. Such a transfer vector preferably comprises a chimeric adenoviral hexon and/or fiber gene sequence as previously described.

Similarly, the means of constructing such a transfer vector are known to those skilled in the art. For instance, a chimeric adenovirus coat protein gene sequence can simply be ligated into the vector using convenient restriction sites. Alternately, a wild-type adenovirus gene sequence can be mutagenized to create the chimeric coat protein sequence following its subcloning into a vector. Similarly, a chimeric coat protein gene sequence

can be moved via standard molecular genetic techniques from a transfer vector into baculovirus or a suitable prokaryotic or eukaryotic expression vector (e.g., a viral or plasmid vector) for expression and evaluation of penton base binding, and other biochemical characteristics.

Accordingly, the present invention also provides recombinant baculoviral and prokaryotic and eukaryotic expression vectors comprising an aforementioned chimeric adenoviral coat protein gene sequence, which, along with the nucleic acid form of the adenoviral vector (i.e., an adenoviral transfer vector) are "transfer vectors" as defined herein. By moving the chimeric gene from an adenoviral vector to baculovirus or a prokaryotic or eukaryotic expression vector, high protein expression is achievable (approximately 5-50% of the total protein being the chimeric protein).

Similarly, adenoviral vectors (e.g., virions or virus particles) are produced using transfer vectors. For instance, an adenoviral vector comprising a chimeric coat protein according to the invention can be constructed by introducing into a cell, e.g., a 293 cell, a vector comprising sequences from the adenoviral left arm, and a vector comprising sequences from the adenoviral right arm, wherein there is a region of overlap between the sequences. As described in the Examples which follow, this methodology results in recombination between the sequences, generating a vector that comprises a portion of each of the vectors, particularly the region comprising the chimeric coat protein sequences.

The present invention thus preferably also provides a method of constructing an adenoviral vector that has a decreased ability or inability to be recognized by a neutralizing antibody directed against wild-type adenovirus hexon protein and/or fiber protein. This method comprises replacing a coat protein of the vector

(i.e., a wild-type adenovirus hexon and/or fiber protein) with the corresponding chimeric adenovirus coat protein according to the invention to produce a recombinant adenoviral vector.

The coat protein chimera-containing particles are produced in standard cell lines, e.g., those currently used for adenoviral vectors. Deletion mutants lacking the fiber gene, or possessing shortened versions of the fiber protein, similarly can be employed in vector construction, e.g., H2d1802, H2d1807, H2d11021 (Falgout et al., supra), as can other fiber mutants. The fiberless particles have been shown to be stable and capable of binding and infecting cells (Falgout et al., supra).

Illustrative Uses and Benefits

The present invention provides a chimeric coat protein that has a decreased ability or inability to be recognized by a neutralizing antibody directed against the corresponding wild-type coat protein, as well as vectors (including transfer vectors) comprising same. The chimeric coat protein (such as a chimeric hexon and/or fiber protein) has multiple uses, e.g., as a tool for studies in vitro of capsid structure and assembly, and capsomere binding to other proteins.

A vector (e.g., a transfer vector) comprising a chimeric coat protein can be used in strain generation, for instance, in generation of recombinant strains of adenovirus. Similarly, such a vector, particularly an adenoviral vector, can be used in gene therapy. Specifically, a vector of the present invention can be used to treat any one of a number of diseases by delivering to targeted cells corrective DNA, i.e., DNA encoding a function that is either absent or impaired, or a discrete killing agent, e.g., DNA encoding a cytotoxin that, for instance, is active only intracellularly.

Diseases that are candidates for such treatment include, but are not limited to, cancer, e.g., melanoma, glioma or lung cancers; genetic disorders, e.g., cystic fibrosis, hemophilia or muscular dystrophy; pathogenic infections, e.g., human immunodeficiency virus, tuberculosis or hepatitis; heart disease, e.g., preventing restenosis following angioplasty or promoting angiogenesis to reperfuse necrotic tissue; and autoimmune disorders, e.g., Crohn's disease, colitis or rheumatoid arthritis. In particular, gene therapy can be carried out in the treatment of diseases, disorders, or conditions that require repeat administration of the corrective DNA and/or the adenoviral vector, and thus for which current adenoviral-mediated approaches to gene therapy are less than optimal.

Moreover, such a vector, particularly an adenoviral vector, can be used to deliver material to a cell not as a method of gene therapy, but for diagnostic or research purposes. In particular, a vector comprising a chimeric adenovirus coat protein according to the invention can be employed to deliver a gene either in vitro or in vivo, for research and/or diagnostic purposes.

For instance, instead of transferring a so-called therapeutic gene, a reporter gene or some type of marker gene can be transferred instead. Marker genes and reporter genes are of use, for instance, in cell differentiation and cell fate studies, as well as potentially for diagnostic purposes. Moreover, a standard reporter gene such as a β -galactosidase reporter gene, a gene encoding green fluorescent protein (GFP), or a β -glucuronidase gene can be used in vivo, e.g., as a means of assay in a living host, or, for instance, as a means of targeted cell ablation (see, e.g., Minden et al., BioTechniques, 20, 122-129 (1996); Youvan, Science, 268,

264 (1995); U.S. Patent 5,432,081; Deonarain et al., <u>Br.</u> J. Cancer, 70, 786-794 (1994)).

Similarly, it may be desirable to transfer a gene to use a host essentially as a means of production in vivo of a particular protein. Along these lines, transgenic animals have been employed, for instance, for the production of recombinant polypeptides in the milk of transgenic bovine species (e.g., PCT International Application WO 93/25567). The use of an adenovirus according to the invention for gene transfer conducted for protein production in vivo further is advantageous in that such use should result in a reduced (if not absent) immune response as compared with the use of a wild-type adenovirus vector. Other "non-therapeutic" reasons for gene transfer include the study of human diseases using an animal model (e.g., use of transgenic mice and other transgenic animals including p53 tumor suppressor gene knockouts for tumorigenic studies, use of a transgenic model for impaired glucose tolerance and human Alzheimer's amyloid precursor protein models for the study of glucose metabolism and for the pathogenesis of Alzheimer's disease, respectively, etc.).

Furthermore, an adenoviral vector comprising a chimeric adenovirus coat protein and employed as described above is advantageous in that it can be isolated and purified by conventional means. For instance, it is likely that special cell lines will not need to be made in order to propagate adenoviruses comprising the chimeric coat proteins.

These aforementioned illustrative uses and recitation of benefits are by no means comprehensive, and it is intended that the present invention encompass such further uses which necessarily flow from, but are not explicitly recited, in the disclosure herein.

Means of Administration

The vectors and transfer vectors of the present invention can be employed to contact cells either in vitro or in vivo. According to the invention "contacting" comprises any means by which a vector is introduced intracellularly; the method is not dependent on any particular means of introduction and is not to be so construed. Means of introduction are well known to those skilled in the art, and also are exemplified herein.

Accordingly, introduction can be effected, for instance, either in vitro (e.g., in an ex vivo type method of gene therapy or in tissue culture studies) or in vivo by methods that include, but are not limited to, electroporation, transformation, transduction, conjugation, triparental mating, (co-)transfection, (co-)infection, high velocity bombardment with DNA-coated microprojectiles, incubation with calcium phosphate-DNA precipitate, direct microinjection into single cells, and the like. Similarly, the vectors can be introduced by means of membrane fusion using cationic lipids, e.g., liposomes. Such liposomes are commercially available (e.g., Lipofectin®, Lipofectamine™, and the like, supplied by Life Technologies, Gibco BRL, Gaithersburg, MD). Moreover, liposomes having increased transfer capacity and/or reduced toxicity in vivo (see, e.g., PCT International Application WO 95/21259 and references reviewed therein) can be employed in the present invention. Other methods also are available and are known to those skilled in the art.

According to the invention, a "host" encompasses any host into which a vector of the invention can be introduced, and thus encompasses an animal, including, but not limited to, an amphibian, bird, insect, reptile, or mammal. Optimally a host is a mammal, for instance, a

rodent, primate (such as chimpanzee, monkey, ape, gorilla, orangutan, or gibbon), feline, canine, ungulate (such as ruminant or swine), as well as, in particular, a human.

Similarly, a "cell" encompasses any cell (or collection of cells) from a host into which an adenoviral vector can be introduced, e.g., preferably an epithelial cell. Any suitable organs or tissues or component cells can be targeted for vector delivery. Preferably, the organs/tissues/cells employed are of the circulatory system (e.g., heart, blood vessels or blood), respiratory system (e.g., nose, pharynx, larynx, trachea, bronchi, bronchioles, lungs), gastrointestinal system (e.g., mouth, pharynx, esophagus, stomach, intestines, salivary glands, pancreas, liver, gallbladder), urinary system (e.g., kidneys, ureters, urinary bladder, urethra), nervous system (e.g. brain and spinal cord, or special sense organs such as the eye) and integumentary system (e.g., skin). Even more preferably the cells being targeted are selected from the group consisting of heart, blood vessel, lung, liver, gallbladder, urinary_bladder, and eye cells.

Thus, the present invention preferably also provides a method of genetically modifying a cell. This method preferably comprises contacting a cell with a vector comprising a chimeric adenovirus hexon protein and/or a chimeric adenovirus fiber protein, wherein desirably the vector is an adenovirus vector. The method preferably results in the production of a host cell comprising a vector according to the invention.

Moreover, the method of the invention of genetically modifying a cell can be employed in gene therapy, or for administration for diagnosis or study. The application of this method in vivo optimally comprises administering to a patient in need of gene therapy (e.g., a patient suffering from a disease, condition or disorder) a therapeutically effective amount of a recombinant adenovirus vector

according to the invention. This method preferably can be employed as part of an ongoing gene therapy regimen, e.g., wherein the vector (e.g., a recombinant adenovirus vector) comprising the chimeric adenovirus coat protein is administered following (e.g., after from about 1 week to about 2 months) administration of a therapeutically effective amount of a vector comprising either the corresponding wild-type coat protein or a coat protein of a different adenoviral serotype. Alternately, the vector comprising the chimeric adenovirus coat protein can be employed as an initial attempt at gene delivery.

One skilled in the art will appreciate that suitable methods of administering a vector (particularly an adenoviral vector) of the present invention to an animal for purposes of gene therapy (see, for example, Rosenfeld et al. (1991), supra; Jaffe et al., Clin. Res., 39(2), 302A (1991); Rosenfeld et al., Clin. Res., 39(2), 311A (1991a); Berkner, supra), chemotherapy, vaccination, diagnosis, and/or further study are available. Although more than one route can be used for administration, a particular route can provide a more immediate and more effective reaction than another route. For instance, local or systemic delivery can be accomplished by administration comprising application or instillation of the formulation into body cavities, inhalation or insufflation of an aerosol, or by parenteral introduction, comprising intramuscular, intravenous, peritoneal, subcutaneous, intradermal, as well as topical administration. Clinical trials regarding use of gene therapy vectors in vivo are ongoing. The methodology employed for such clinical trials as well as further technologies known to those skilled in the art can be used to administer the vector of the present invention for the purpose of research, diagnosis and/or gene therapy.

Pharmaceutically acceptable excipients also are well-known to those who are skilled in the art, and are readily available. The choice of excipient will be determined in part by the particular method used to administer the recombinant vector. Accordingly, there is a wide variety of suitable formulations for use in the context of the present invention. The following methods and excipients are merely exemplary and are in no way limiting.

Formulations suitable for oral administration can consist of (a) liquid solutions, such as an effective amount of the compound dissolved in diluents, such as water, saline, or orange juice; (b) capsules, sachets or tablets, each containing a predetermined amount of the active ingredient, as solids or granules; (c) suspensions in an appropriate liquid; and (d) suitable emulsions. Tablet forms can include one or more of lactose, mannitol, corn starch, potato starch, microcrystalline cellulose, acacia, gelatin, colloidal silicon dioxide, croscarmellose sodium, talc, magnesium stearate, stearic acid, and other excipients, colorants, diluents, buffering agents, moistening agents, preservatives, flavoring agents, and pharmacologically compatible excipients. Lozenge forms can comprise the active ingredient in a flavor, usually sucrose and acacia or tragacanth, as well as pastilles comprising the active ingredient in an inert base, such as gelatin and glycerin, or sucrose and acacia, emulsions, gels, and the like containing, in addition to the active ingredient, such excipients as are known in the art.

A vector of the present invention (including an adenoviral vector and a transfer vector), alone or in combination with other suitable components, can be made into aerosol formulations to be administered via inhalation. These aerosol formulations can be placed into pressurized acceptable propellants, such as dichlorodifluoromethane, propane, nitrogen, and the like.

They may also be formulated as pharmaceuticals for non-pressured preparations such as in a nebulizer or an atomizer.

Formulations suitable for parenteral administration include aqueous and non-aqueous, isotonic sterile injection solutions, which can contain anti-oxidants, buffers, bacteriostats, and solutes that render the formulation isotonic with the blood of the intended recipient, and aqueous and non-aqueous sterile suspensions that can include suspending agents, solubilizers, thickening agents, stabilizers, and preservatives. formulations can be presented in unit-dose or multi-dose sealed containers, such as ampules and vials, and can be stored in a freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid excipient, for example, water, for injections, immediately prior to use. Extemporaneous injection solutions and suspensions can be prepared from sterile powders, granules, and tablets of the kind previously described.

Additionally, a vector of the present invention can be made into suppositories by mixing with a variety of bases such as emulsifying bases or water-soluble bases.

Formulations suitable for vaginal administration can be presented as pessaries, tampons, creams, gels, pastes, foams, or spray formulas containing, in addition to the active ingredient, such carriers as are known in the art to be appropriate.

The dose administered to an animal, particularly a human, in the context of the present invention will vary with the gene of interest, the composition employed, the method of administration, the particular site and organism undergoing administration, and the reason for the administration (e.g., gene therapy, diagnosis, means of producing a protein, further study, etc). Generally, the "effective amount" of the composition is such as to

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produce the desired effect in a host which can be monitored using several end-points known to those skilled in the art. For example, one desired effect might comprise effective nucleic acid transfer to a host cell. Such transfer can be monitored in terms of a therapeutic effect (e.g., alleviation of some symptom associated with the disease or syndrome being treated), or by further evidence of the transferred gene or coding sequence or its expression within the host (e.g., using the polymerase chain reaction, Northern or Southern hybridizations, or transcription assays to detect the nucleic acid in host cells, or using immunoblot analysis, antibody-mediated detection, or particularized assays to detect protein or polypeptide encoded by the transferred nucleic acid, or impacted in level or function due to such transfer). One such particularized assay described in the Examples which follow includes an assay for expression of a chloramphenicol acetyl transferase reporter gene.

Generally, to ensure effective transfer of the vectors of the present invention, it is preferable that from about 1 to about 5,000 copies of the vector be employed per cell to be contacted, based on an approximate number of cells to be contacted in view of the given route of administration. It is even more preferable that from about 1 to about 300 plaque forming units (pfu) enter each cell. However, this is just a general guideline which by no means precludes use of a higher or lower amount of a component, as might be warranted in a particular application, either in vitro or in vivo. For example, the actual dose and schedule can vary depending on whether the composition is administered in combination with other pharmaceutical compositions, or depending on interindividual differences in pharmacokinetics, drug disposition, and metabolism. Similarly, amounts can vary in in vitro applications depending on the particular cell

type utilized or the means by which the vector is transferred. One skilled in the art easily can make any necessary adjustments in accordance with the necessities of the particular situation.

The following examples further illustrate the present invention and, of course, should not be construed as in any way limiting its scope.

Example 1

This example describes experiments investigating adenoviral anti-vector neutralizing immunity.

To clarify the phenomenon of neutralizing immunity, an animal having circulating antibodies to one adenoviral vector type received intratracheal administration of another serotype adenoviral vector, and gene expression commanded by the second vector was monitored.

Specifically, either an Ad4 or Ad5 wild-type vector was administered to the lungs of Sprague-Dawley rats. Ten days later, an Ad5 reporter vector was administered to the lungs of the same animals. This reporter vector, which is referred to herein as the "pure 5" vector, comprises an E1-E3-type 5 adenoviral vector which expresses the chloramphenical acetyl transferase (CAT) gene driven by the cytomegalovirus early/intermediate promoter/enhancer (CMV) (i.e., AdCMVCATgD described in Kass-Eisler et al., Proc. Natl. Acad. Sci., 15, 11498-11502 (1993)).

About twenty-four hours following administration of the "pure 5" vector, CAT activity was measured in homogenized lung tissue using a CAT assay as previously described (Kass Eisler et al. (1993), supra). CAT activity was monitored at various times thereafter up to 10 days following introduction of the "pure 5" vector. CAT activity was determined relative to the "pure 5" vector administered to naive animals (i.e., expression measured under this condition was considered 100%). The

results of these studies are set out in **Table 1**, and are further reported in Mastrangeli et al., <u>Human Gene</u>
Therapy, 1, 79-87 (1996).

Table 1. Effect of anti-serotype 4 (group E) neutralizing antibodies on the ability of a "pure 5"
adenoviral vector to transfer a CAT reporter gene to
the lung

Time (0 hours)	Time (10 days)	CAT Activity
		0%
	pure 5	100%
Ad5	pure 5	. 0%
Ad4	pure 5	105±10%

These results confirm that in the presence of neutralizing antibodies elicited against one adenoviral group (e.g., against group E, serotype 4), it is possible to efficiently transfer and express a gene in vivo using an adenoviral vector derived from another group (e.g., derived from group C, serotype 5). Neutralizing immunity evoked against one serotype group does not protect against infection by another group of adenovirus. These data support the paradigm of alternating adenoviral vectors derived from different subgroups as a strategy to circumvent anti-adenoviral humoral immunity.

Example 2

The predominant epitopes that evoke neutralizing immunity are located on the fiber and hexon, but mainly on hexon. Based on this, the effect of switching the fiber protein was investigated. A vector was constructed that was identical to the "pure 5" vector except that the fiber gene was switched from a serotype 5, group C fiber to a

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serotype 7, group B fiber. The resultant vector is referred to herein as the "5 base/7 fiber" vector.

The Ad5/Ad7 fiber construct was generated as shown in Figure 1. An approximately 2.7 kb (Ad5 28689-31317 bp) fragment in pAd70-100 was replaced with a PacI linker (pAd70-100dlE3.Pac). A BamHI linker was inserted at a MunI site as indicated in Figure 2 to produce pAd70-100dlE3.Pac.Bam. A PCR-amplified PacI-BamHI fragment of approximately 1.1 kb containing the Ad7 fiber gene was inserted into pAd70-100dlE3.Pac.Bam to produce pAd70-100dlE3.fiber7.

In order to assess the ability of the Ad5 virus with Ad7 fiber to infect cells in vitro and in vivo, reporter gene assays were performed. A replication-defective recombinant adenoviral reporter vector designated AdCMV-CATNeo was used in the reporter gene assay. The reporter vector consists of the adenoviral origin of replication and viral packaging sequences, a combination of strong eukaryotic promoter (cytomegalovirus or CMV-1) and splicing elements, the bacterial chloramphenicol acetyl transferase (CAT) gene sequence, the mouse $\beta^{\text{maj}}\text{-globin}$ poly(A) site, the neomycin gene sequence (Neo), and sufficient adenoviral DNA to allow for overlap recombination.

The reporter vector was used to generate AdCMV-CATNeo, AdCMV-CATNeo-dlE3 (AdCMV-CATNeo + pAd70-100dlE3) and AdCMV-CATNeo-dlE3-Fiber7 (AdCMV-CATNeo + pAd70-1001E3.Fiber7) viruses. Each virus was grown in large scale, i.e., a one liter suspension of human embryonic kidney 293 cells, to yield virus at a concentration of 10¹² particles/ml. A549 cells were infected with an estimated 100, 300 or 1,000 particles/cell of one of the three viruses. After 48 hours, the cells were harvested and lysates were prepared as described in Kass-Eisler et al.

(1993), supra. Using 50 μ l of each lysate, CAT assays were performed and acetylated chloramphenical products were separated by thin layer chromatography using chloroform:methanol (95:5). The results of the assays confirm that each virus was able to infect cells and express gene products at appropriate levels. Accordingly, the virus in which the native fiber was replaced with a nonnative fiber could infect cells and express genes like the parental virus.

Following this study, adult Sprague-Dawley rats were infected with 108 viral particles by direct cardiac injection as described in Kass-Eisler et al. (1993), supra. Five days later, the rats were sacrificed, cardiac lysates were prepared, and CAT assays were performed. amount of the CAT gene product produced was compared between the dlE3 and dlE3-Fiber7 viruses. Results indicated that both viruses were able to infect cells in vivo. The replacement of the wild-type Ad5 fiber gene with that of Ad7 did not impair the ability of the virus to infect cells. Accordingly, the virus in which the native fiber was replaced with a nonnative fiber could also infect cells and express genes like the parental virus in vivo. These results support the utility of adenovirus with chimeric fiber in the context of gene therapy.

Example 3

This example describes the effect on neutralizing immunity of switching the fiber protein of an adenovirus from one serotype to another.

The "pure 5" and "5 base/7 fiber" vectors described in the preceding Example were administered to Sprague-Dawley rats which either were naive or pre-immunized against wild-type Ad5. For these experiments, wild-type Ad5 or wild-type Ad7 (6 x 109 particles in phosphate

buffered saline (PBS)) was administered intraperitoneally as a primary inoculation. Seventeen days later, serum samples were taken, and about 6 x 10° particles in about 50 µl of PBS was injected. At about 120 hours following injection the animals were sacrificed, serum and heart tissue were harvested, and heart tissue was processed for CAT assays as previously described (Kass-Eisler et al. (1993), supra). CAT assays also were performed on heart lysates of rat hearts infected with the "pure 5" vector or "5 base/7 fiber" vector alone.

Administration of either vector to naive animals resulted in comparable levels of CAT in heart tissue. In comparison, administration of either the "pure 5" vector or the "5 base/7 fiber" vector to the animals that were pre-immunized against the "pure 5" vector resulted in a reduction of CAT levels by more than two orders of magnitude as compared with mock-infected controls. These and further results are reported in Gall et al., <u>J. Virol.</u>, <u>70</u>, 2116-2163 (1996).

These results confirm that switching the fiber from that of adenoviral serotype 5 group C vector to that of an adenoviral serotype 7 group B vector by itself is insufficient to allow the vector to escape neutralizing antibodies generated against an adenoviral vector comprising Ad5 fiber. These results imply that antibodies against adenoviral structures other than fiber also are important in the process of neutralizing immunity. Furthermore, whereas switching the fiber serotype to another serotype may be insufficient in and of itself to allow an adenovirus to escape immune detection, such switching when done in combination with removal of other epitopes may be desirable, for instance, to reduce an immune response.

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Example 4

This example describes the construction of adenovirus vectors wherein the neutralizing immunity-evoking epitopes have been modified. In particular, this example describes vectors comprising chimeric adenoviral hexon protein, wherein the hexon neutralizing immunity-evoking epitopes are modified.

The results of the prior example indicate that it is possible to develop vectors for repeat administration in gene therapy from non-group C adenovirus, thus circumventing pre-existing neutralizing immunity. As another strategy, the dominant neutralizing immunity-evoking epitopes on existing group C vectors can be modified to render the vectors less susceptible (or "stealth") to the existing neutralizing immunity. For instance, adenoviral type 5-based E1 E3 CAT-expressing vectors can be constructed that have the same genetic composition as the "pure 5" and "5 base/7 fiber" vectors described above, except for possessing a gene encoding a chimeric hexon that is not recognized by pre-existing anti-type 5 neutralizing immunity.

To derive the vectors, the chimeric hexon gene present in the "pure 5" parental vector can be modified, in particular, 11 and/or 12 can be altered. The hexon modifications that can be made on the "pure 5" CAT vector, or other adenoviral vector (such as any other adenoviral serotype vector), include, but are not limited to: (1) hexon with 11 deleted in its entirety; (2) hexon with 12 deleted in its entirety; (3) hexon with both 11 and 12 deleted; (4) hexon with any one or more of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7, deleted; (5)-(8) hexon with a FLAG octamer epitope (i.e., Asp Tyr Lys Asp Asp Asp Asp Lys [SEQ ID NO:50]; Hopp et al., Biotechnology, 6, 1205-1210 (1988)) substituted for 11, 12, or both 11 and 12, or any one or more of HVR1, HVR2, HVR3, HVR4, HVR5,

HVR6 or HVR7; (9)-(12) hexon with a FLAG octamer epitope [SEQ ID NO:50] inserted into 11, 12, or both 11 and 12; (13)-(16) hexon with comparable epitopes from Ad7 (group B) (GenBank® Data Bank Accession Number x76551 for Ad7 hexon, and Number M73260 for Ad5 hexon) or Ad2, or any other adenoviral serotype, substituted for 11, 12, both 11 and 12, respectively, or for any one or more of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7; (17)-(20) hexon with comparable epitopes from Ad7 (group B) (GenBank® Data Bank Accession Number x76551 for Ad7 hexon, and Number M73260 for Ad5 hexon) or Ad2, or any other adenoviral serotype, inserted into 11, 12, both 11 and 12, respectively, or any one or more of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7; and (21) complete substitution of the hexon from Ad2 or another adenoviral serotype, for the Ad5 hexon. The use of the FLAG octamer epitope provides a sequence for incorporation in the chimeric hexon protein that is different from the Ad5 hexon loop sequences, and also provides a positive control using available specific anti-FLAG antibodies (Hopp et al., supra).

These chimeric hexon proteins (and vectors containing them) can be made in several steps. To modify the hexon in the "pure 5" vector, a viral or plasmid vector can be constructed to contain the hexon type 5 coding sequence in a cassette that can be easily modified. The hexon is read off the 1 strand of the L3 transcription unit, i.e., map units 51.6 to 59.7, comprising a region of about 2.9 kb. The two other transcripts that also are encoded by L3 — i.e., polypeptide VI and a 23 kDa protein — do not overlap the hexon coding sequence. Moreover, there are no other coding sequences on the r strand that would be altered by the modification of the hexon coding sequence.

Thus, all the modifications of the type 5 hexon can be made using a "hexon 5 cassette" comprised of an

approximate 6.7 kb <u>SfiI-SfiI</u> fragment of the "pure 5" CAT vector. *SfiI* cuts Ad5 into 3 fragments, the center 6.7 kb fragment (i.e., comprising about 16,282 to 22,992 base pairs, as identified by agarose gel electrophoresis) of which contains all of the L3 region plus some overlap. The "hexon 5 cassette" can be subcloned into a commercially available vector having restriction sites and the like making the vector easily manipulable in terms of modification and recovery of subcloned sequences. One such vector appropriate for subcloning is either the SK or KS version of the pBlueScript® phagemid (Stratagene, LaJolla, CA).

The "hexon 5 cassette" can be mutagenized to generate site-specific mutations in the cloned DNA segment. Several methods are available for carrying out sitespecific mutagenesis. The 11 and 12 deletions, insertions, or replacements (or deletions, insertions, or replacements in HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7 regions contained therein) can be made by deleting the relevant sequences using restriction enzymes that cut uniquely within the vector inserts, or other similar means, e.g., by ligating in an end-polished, or otherwise modified, PCR product. Alternately, the hexon sequence contained in the hexon 5 cassette can be modified, e.g., using single-stranded mutagenesis in M13mp8 or some other convenient vector, and using appropriate oligonucleotides encompassing the flanking sequences for identification of plaques as described by Crompton et al., supra. Alternately, a commercially available kit such as the ExSiteTM PCR-based site-directed mutagenesis kit and the Chameleon™ double-stranded site-directed mutagenesis kit by Stratagene can be used to introduce insertions, point mutations, or deletions into the chimeric hexon sequence without any need for subcloning into an M13, or other special vector.

Similarly, the FLAG octapeptide sequence (Hopp et al., supra) can be introduced into the vectors (i.e., in the presence or absence of any deletion) by inserting the relevant 24 base pair sequence (GAY TAY AAR GAY GAY GAY GAY AAR [SEQ ID NO:50], wherein Y is C or T/U, and R is A or G)). The replacement of Ad5 hexon loop epitopes with comparable sequences of Ad7, Ad2, or any other adenoviral serotype, or an incorporation of these sequences in the absence of any deletion, can be accomplished by using unique restriction sites, or using one of the aforementioned means of mutagenesis. This usefully creates new serotypes of adenoviral vectors. For example, The replacement of the wildtype hexon protein of Ad5 with the chimeric Ad5 hexon comprising Ad7 hexon loops 1 and 2 gives rise to an adenoviral vector that is effectively neutralized by Ad7 neutralizing antibodies (i.e., neutralizing antibodies raised in response to Ad7 innoculation of a naïve animal), but not by Ad5 neutralizing antibodies.

Moreover, both hypervariable loops 1 and 2 can be deleted from a serotype 5 or another serotype adenoviral vector. Adenoviral vectors and there genomes comprising these deletions are useful as a starting point to create other adenoviral vectors having loop replacements, as a tool for studying hexon structure-function relationships, and under some circumstances as a gene transfer vector with limited vulnerability to the adaptive immune system.

Example 5

This example describes the method of replacing the hexon protein of one serotype adenoviral vector with the hexon protein of another serotype adenoviral vector to generate a recombinant adenovirus. As representative of this method, the hexon protein of an Ad5 vector was replaced with the hexon protein of an Ad2 vector. This

example also describes the method of incorporating the chimeric hexon proteins of the preceding Example into a vector to make a recombinant adenovirus.

Using standard molecular biology techniques, the Ad5 hexon gene open reading frame (ORF) was replaced with the Ad2 hexon gene ORF in such a fashion so as to maintain the proper Ad5 sequences upstream and downstream of the hexon gene. Adenoviral vectors comprising modified or chimeric hexon proteins can be constructed by homologous recombination using standard techniques and human embryonic kidney 293 cells (see, e.g., Rosenfeld et al. (1991), supra; Rosenfeld et al. (1992), supra). For instance, map units 0 to 57.3 of dlAd5NCAT (Gall et al., supra) can be isolated by Bsu36I digestion, and map units 58.4 to 100 of dlAd5NCAT can be isolated by DrdI digestion. These DNA fragments can be transfected into 293 cells along with pH5-2.

A neutralizing antibody directed against the parental vector can be employed to facilitate the generation of hexon replacement constructs. For example, when replacing the loop 1 and loop 2 regions of an Ad5 vector with Ad7 loop sequences, anti-Ad5 neutralizing polyclonal or monoclonal antibodies (directed against the loops 1 and 2 of Ad5 hexon) can be added to a the medium of cells in which the chimeric vector is being propagated. presence of the Ad5 neutralizing antibodies substantially blocks the propagation of the undesired wildtype Ad5 vector(s), while the chimeric vector is unaffected. Furthermore, the recombinant vectors comprising a chimeric hexon ORF can be generated by homologous recombination using a plasmid that carries a marker gene, such as Green Fluorescent Protein (GFP), adjacent to the chimeric or novel hexon ORF (e.g., between the fiber and hexon genes). In this way, genomes that could harbor the chimeric hexon gene should also harbor the marker gene. The marker gene

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would then be expressed as a late protein, so that cells that potentially comprise the desired adenoviral genome can be easily identified.

Similarly, vectors (particularly adenoviral vectors) can be constructed that have the aforementioned hexon modifications, and which have further modifications, for instance, in the adenoviral fiber coding sequences. This can be accomplished by making the hexon modifications described above, and using different parental plasmids for homologous recombination, such as parental plasmids comprising mutations in fiber coding sequences. In particular, the "5 base/7 fiber" vector can be employed as a starting vector for vector construction.

All of the viral vectors prepared according to this example can be plaque-purified, amplified, and further purified using standard methods (Rosenfeld et al. (1991), supra; Rosenfeld et al. (1992), supra).

Example 6

This example describes a characterization of the activity in vitro and in vivo of the vectors described in the preceding Examples.

Each of the viruses prepared as described in the preceding Examples can be evaluated in vitro and in vivo using standard methods as previously described (e.g., Kass-Eisler et al., supra), and as set forth herein. In particular, for the in vitro studies, the various vectors along with control vectors (e.g., the "pure 5" and "5 base/7 fiber" vectors, and the Ad5 wild-type vector) can be added to human lung carcinoma A549 cells alone, or in the presence of dilutions of serum from hosts infected with Ad5, Ad7, "pure 5" CAT vector, or "5 base/7 fiber" CAT vector, or anti-FLAG epitope serum. The cells are then evaluated for CAT activity to determine the ability

of antibodies present in the serum to block gene expression.

The in vivo studies can be carried out in Sprague-Dawley rats. The Sprague-Dawley rat as opposed to the mouse or cotton rat is preferred for these experiments since the rat is non-permissive, and the wild-type adenovirus cannot replicate in this host. Accordingly, immunizations can be carried out using wild-type viruses (e.g., wild-type Ad5 or Ad7), the "pure 5" CAT vector, and the "5 base/7 fiber" CAT vector by intravenous administration (e.g., Kass-Eisler et al., supra). various times ranging from about one to about four weeks later, the vector of interest can be administered intravenously or directly into the airways of the host. Whereas intravenous administration allows an assessment of the "worst case scenario" (i.e., wherein the vector is in immediate contact with the circulating humoral immune system, and thus the strongest immune response is to be expected), introduction in the airways of the host allows an evaluation of a compartmentalized and mucosal humoral immune response.

CAT activity can be quantified as previously described in all the relevant organs, e.g., liver, heart, and lung for intravenous administration, and lung only for respiratory administration. Appropriate standards can be used to compensate for variations in organ expression of CAT activity (see e.g., Kass-Eisler et al., Gene Therapy, 2 395-402 (1994)). The in vitro and in vivo results can be compared and assessed using standard statistical methods.

All of the references cited herein, including the GenBank® Data Bank sequence information, are hereby incorporated in their entireties by reference.

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While this invention has been described with emphasis upon preferred embodiments, it will be obvious to those of ordinary skill in the art that the preferred embodiments can be varied. It is intended that the invention can be practiced otherwise than as specifically described herein. Accordingly, this invention includes all modifications encompassed within the spirit and scope of the appended claims.

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- (ii) TITLE OF INVENTION: CHIMERIC ADENOVIRAL COAT PROTEIN AND METHODS OF USING SAME
- (iii) NUMBER OF SEQUENCES: 56
- (iv) COMPUTER READABLE FORM:

		(E	3) CC	MPUT ERAT	ER:	IBM SYST	PC c	PC-E	tib]	1S-DC	/ersi	ion #	1.30) (EP	D)
	(vi)	PRI (F	OR A	PPLIC	CATI	ON E	ATA: IMBER	:	8-8						
(2)	INE	ORMA	OITA	FOF	SEC] ID	NO: 1	l:							
	(i)	(F (E	QUENCA) LE B) TY C) SI C) TO	NGTH PE: RANE	l: 29 nucl EDNE	007 b eic ESS:	ase acio doub	pair 1	:s					•	
	(ii)	MOI	ECUI	E TY	PE:	DNA	(ger	nomic	=)		•				
	(xi)	SEÇ	QUENC	CE DE	SCRI	PTIC)N: 5	SEQ 1	D NO):1:	•	•			
	Ala				Met					Ser		CAC His		Ser	48
												CAG Gln 30			96
												AGA Arg			144
												CAG Gln			192
												TAC Tyr			240
												CTT Leu			288
												GGG Gly 110			336
												CCC Pro			384
												GGC Gly			432
												GAG Glu		GAA Glu 160	480
												CAT His			528

					TCT Ser									576
					GCA Ala									624
					GAA Glu									672
					GCA Ala 230									720
					GGA Gly									768
	_		_		GTT Val									816
					TTC Phe									864
					CCA Pro									912
					ACA Thr 310									960
					ATG Met									1008
												AAC Asn		1056
					GGT Gly		Ala						•	1104
		Val										CTC Leu		1152
												AAT Asn 400		1200
					Tyr				Arg			CAT His	•	1248
	-			Glu				Cys				ATT Ile		1296

					TAT Tyr											1344
					ACT Thr											1392
					GTG Val 470											1440
					AGA Arg											1488
					AAA Lys											1536
AAC Asn	CCC Pro	AAC Asn 515	ACC Thr	TAC Tyr	GAC Asp	TAC Tyr	ATG Met 520	AAC Asn	AAG Lys	CGA Arg	GTG Val	GTG Val 525	GCT Ala	CCC Pro	GGG Gly	1584
					ATT											1632
					CCC Pro 550											1680
TAT Tyr	CGC Arg	TCC Ser	ATG Met	TTG Leu 565	TTG Leu	GGA Gly	AAC Asn	GGC Gly	CGC Arg 570	TAC Tyr	GTG Val	CCC Pro	TTT Phe	CAC His 575	ATT Ile	1728
CAG Gln	GTG Val	CCC Pro	CAA Gln 580	AAG Lys	TTT Phe	TTT Phe	GCC Ala	ATT Ile 585	AAA Lys	AAC Asn	CTC Leu	CTC Leu	CTC Leu 590	CTG Leu	CCA Pro	1776
GGC Gly	TCA Ser	TAT Tyr 595	ACA Thr	TAT Tyr	GAA Glu	TGG Trp	AAC Asn 600	TTC Phe	AGG Arg	AAG Lys	GAT Asp	GTT Val 605	AAC Asn	ATG Met	GTT Val	1824
					GGA Gly											1872
					TGT Cys 630											1920
AAC Asn	ACG Thr	GCC Ala	TCC Ser	ACG Thr 645	CTG Leu	GAA Glu	GCC Ala	ATG Met	CTC Leu 650	AGA Arg	TAA Asn	GAC Asp	ACC Thr	AAC Asn 655	GAC Asp	1968
					TAC Tyr											2016
CCC Pro	GCC Ala	AAC Asn 675	GCC Ala	ACC Thr	AAC Asn	GTG Val	CCC Pro 680	ATC Ile	TCC Ser	ATC Ile	CCA Pro	TCG Ser 685	CGC Arg	AAC Asn	TGG Trp	2064

GCA Ala	GCA Ala 690	TTT Phe	CGC Arg	GGT Gly	Trp	GCC Ala 695	TTC Phe	ACA Thr	CGC Arg	TTG Leu	AAG Lys 700	ACA Thr	AAG Lys	GAA Glu	ACC Thr	2112
CCT Pro 705	TCC Ser	CTG Leu	GGA Gly	TCA Ser	GGC Gly 710	TAC Tyr	GAC Asp	CCT Pro	TAC Tyr	TAC Tyr 715	ACC Thr	TAC Tyr	TCT Ser	GGC Gly	TCC Ser 720	2160
ATA Ile	CCA Pro	TAC Tyr	CTT Leu	GAC Asp 725	GGA Gly	ACC Thr	TTC Phe	TAT Tyr	CTT. Leu 730	AAT Asn	CAC His	ACC Thr	TTT Phe	AAG Lys 735	AAG Lys	2208
GTG Val	GCC Ala	ATT Ile	ACC Thr 740	TTT Phe	GAC Asp	TCT Ser	TCT Ser	GTT Val 745	AGC Ser	TGG Trp	CCG Pro	GGC Gly	AAC Asn 750	GAC Asp	CGC Arg	2256
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GGC Gly	TAC Tyr 770	AAC Asn	GTA Val	GCT Ala	CAG Gln	TGC Cys 775	AAC Asn	ATG Met	ACC Thr	AAG Lys	GAC Asp 780	TGG Trp	TTC Phe	CTG Leu	GTG Val	2352
CAG Gln 785	ATG Met	TTG Leu	GCC Ala	AAC Asn	TAC Tyr 790	AAT Asn	ATT Ile	GGC Gly	TAC Tyr	CAG Gln 795	GGC Gly	TTC Phe	TAC Tyr	ATT Ile	CCA Pro 800	2400
GAA Glu	AGC Ser	TAC Tyr	AAG Lys	GAC Asp 805	CGC Arg	ATG Met	TAC Tyr	TCG Ser	TTC Phe 810	TTC Phe	AGA Arg	AAC Asn	TTC Phe	CAG Gln 815	CCC Pro	2448
ATG Met	AGC Ser	CGG Arg	CAA Gln 820	GTG Val	GTT Val	GAC Asp	GAT Asp	ACT Thr 825	AAA Lys	TAC Tyr	AAG Lys	GAG Glu	TAT Tyr 830	CAG Gln	CAG Gln	2496
GTT Val	GGA Gly	ATT Ile 835	CTT Leu	CAC His	CAG Gln	CAT His	AAC Asn 840	AAC Asn	TCA Ser	GGA Gly	TTC Phe	GTA Val 845	GGC Gly	TAC Tyr	CTC Leu	2544
GCT Ala	CCC Pro 850	Thr	ATG Met	CGC Arg	GAG Glu	GGA Gly 855	CAG Gln	GCT Ala	TAC Tyr	CCC Pro	GCC Ala 860	AAC Asn	GTG Val	CCC Pro	TAC Tyr	2592
CCA Pro 865	Leu	ATA Ile	GGC Gly	AAA Lys	ACC Thr 870	GCG Ala	GTT Val	GAC Asp	AGT Ser	ATT Ile 875	Thr	CAG Gln	AAA Lys	AAG Lys	TTT Phe 880	2640
CTT Leu	TGC	GAT Asp	CGC Arg	ACC Thr 885	Leu	TGG Trp	CGC Arg	ATC Ile	CCA Pro 890	Phe	TCC Ser	AGT Ser	AAC Asn	TTT Phe 895	ATG Met	2688
TCC Ser	ATG Met	GGC	GCA Ala 900	Leu	ACA Thr	GAC Asp	CTG Leu	GGC Gly 905	Gln	AAC Asn	CTT Leu	CTC Leu	TAC Tyr 910	Ala	AAC Asn	2736
TCC Ser	GCC Ala	CAC His 915	Ala	CTA Leu	GAC Asp	ATG Met	ACT Thr 920	Phe	GAG Glu	GTG Val	GAT Asp	CCC Pro 925	Met	GAC Asp	GAG Glu	2784
CCC	ACC Thr 930	Leu	CTT Leu	TAT	GTT Val	TTG Leu 935	Phe	GAA Glu	GTC Val	TTT Phe	GAC Asp 940	Val	GTC Val	CGT Arg	GTG Val	2832

2880 CAC CAG CCG CAC CGC GGC GTC ATC GAG ACC GTG TAC CTG CGC ACG CCC His Gln Pro His Arg Gly Val Ile Glu Thr Val Tyr Leu Arg Thr Pro 950 2907 TTC TCG GCC GGC AAC GCC ACA ACA TAA Phe Ser Ala Gly Asn Ala Thr Thr 965 (2) INFORMATION FOR SEQ ID NO:2: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 968 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2: Met Ala Thr Pro Ser Met Met Pro Gln Trp Ser Tyr Met His Ile Ser Gly Gln Asp Ala Ser Glu Tyr Leu Ser Pro Gly Leu Val Gln Phe Ala Arg Ala Thr Glu Thr Tyr Phe Ser Leu Asn Asn Lys Phe Arg Asn Pro Thr Val Ala Pro Thr His Asp Val Thr Thr Asp Arg Ser Gln Arg Leu Thr Leu Arg Phe Ile Pro Val Asp Arg Glu Asp Thr Ala Tyr Ser Tyr Lys Ala Arg Phe Thr Leu Ala Val Gly Asp Asn Arg Val Leu Asp Met Ala Ser Thr Tyr Phe Asp Ile Arg Gly Val Leu Asp Arg Gly Pro Thr Phe Lys Pro Tyr Ser Gly Thr Ala Tyr Asn Ala Leu Ala Pro Lys Gly Ala Pro Asn Ser Cys Glu Trp Glu Gln Thr Glu Asp Ser Gly Arg Ala Val Ala Glu Asp Glu Glu Glu Glu Asp Glu Glu Glu Glu Glu Glu Glu Glu Gln Asn Ala Arg Asp Gln Ala Thr Lys Lys Thr His Val Tyr Ala Gln Ala Pro Leu Ser Gly Glu Thr Ile Thr Lys Ser Gly Leu Gln Ile Gly Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val Tyr Ala Asp Pro Ser Tyr Gln Pro Glu Pro Gln Ile Gly Glu Ser Gln Trp Asn Glu

Ala Asp Ala Asn Ala Ala Gly Gly Arg Val Leu Lys Lys Thr Thr Pro

Met Lys Pro Cys Tyr Gly Ser Tyr Ala Arg Pro Thr Asn Pro Phe Gly Gly Gln Ser Val Leu Val Pro Asp Glu Lys Gly Val Pro Leu Pro Lys Val Asp Leu Gln Phe Phe Ser Asn Thr Thr Ser Leu Asn Asp Arg Gln Gly Asn Ala Thr Lys Pro Lys Val Val Leu Tyr Ser Glu Asp Val Asn Met Glu Thr Pro Asp Thr His Leu Ser Tyr Lys Pro Gly Lys Gly Asp Glu Asn Ser Lys Ala Met Leu Gly Gln Gln Ser Met Pro Asn Arg Pro 330 Asn Tyr Ile Ala Phe Arg Asp Asn Phe Ile Gly Leu Met Tyr Tyr Asn Ser Thr Gly Asn Met Gly Val Leu Ala Gly Gln Ala Ser Gln Leu Asn Ala Val Val Asp Leu Gln Asp Arg Asn Thr Glu Leu Ser Tyr Gln Leu Leu Leu Asp Ser Ile Gly Asp Arg Thr Arg Tyr Phe Ser Met Trp Asn Gln Ala Val Asp Ser Tyr Asp Pro Asp Val Arg Ile Ile Glu Asn His Gly Thr Glu Asp Glu Leu Pro Asn Tyr Cys Phe Pro Leu Gly Gly Ile Gly Val Thr Asp Thr Tyr Gln Ala Ile Lys Ala Asn Gly Asn Gly Ser Gly Asp Asn Gly Asp Thr Thr Trp Thr Lys Asp Glu Thr Phe Ala Thr 455 Arg Asn Glu Ile Gly Val Gly Asn Asn Phe Ala Met Glu Ile Asn Leu Asn Ala Asn Leu Trp Arg Asn Phe Leu Tyr Ser Asn Ile Ala Leu Tyr Leu Pro Asp Lys Leu Lys Tyr Asn Pro Thr Asn Val Glu Ile Ser Asp Asn Pro Asn Thr Tyr Asp Tyr Met Asn Lys Arg Val Val Ala Pro Gly Leu Val Asp Cys Tyr Ile Asn Leu Gly Ala Arg Trp Ser Leu Asp Tyr Met Asp Asn Val Asn Pro Phe Asn His His Arg Asn Ala Gly Leu Arg 550 Tyr Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile

Gln Val Pro Gln Lys Phe Phe Ala Ile Lys Asn Leu Leu Leu Pro Gly Ser Tyr Thr Tyr Glu Trp Asn Phe Arg Lys Asp Val Asn Met Val 600 Leu Gln Ser Ser Leu Gly Asn Asp Leu Arg Val Asp Gly Ala Ser Ile Lys Phe Asp Ser Ile Cys Leu Tyr Ala Thr Phe Phe Pro Met Ala His Asn Thr Ala Ser Thr Leu Glu Ala Met Leu Arg Asn Asp Thr Asn Asp 650 Gln Ser Phe Asn Asp Tyr Leu Ser Ala Ala Asn Met Leu Tyr Pro Ile Pro Ala Asn Ala Thr Asn Val Pro Ile Ser Ile Pro Ser Arg Asn Trp Ala Ala Phe Arg Gly Trp Ala Phe Thr Arg Leu Lys Thr Lys Glu Thr Pro Ser Leu Gly Ser Gly Tyr Asp Pro Tyr Tyr Thr Tyr Ser Gly Ser Ile Pro Tyr Leu Asp Gly Thr Phe Tyr Leu Asn His Thr Phe Lys Lys Val Ala Ile Thr Phe Asp Ser Ser Val Ser Trp Pro Gly Asn Asp Arg Leu Leu Thr Pro Asn Glu Phe Glu Ile Lys Arg Ser Val Asp Gly Glu Gly Tyr Asn Val Ala Gln Cys Asn Met Thr Lys Asp Trp Phe Leu Val 770 780 Gln Met Leu Ala Asn Tyr Asn Ile Gly Tyr Gln Gly Phe Tyr Ile Pro Glu Ser Tyr Lys Asp Arg Met Tyr Ser Phe Phe Arg Asn Phe Gln Pro 810 Met Ser Arg Gln Val Val Asp Asp Thr Lys Tyr Lys Glu Tyr Gln Gln Val Gly Ile Leu His Gln His Asn Asn Ser Gly Phe Val Gly Tyr Leu Ala Pro Thr Met Arg Glu Gly Gln Ala Tyr Pro Ala Asn Val Pro Tyr Pro Leu Ile Gly Lys Thr Ala Val Asp Ser Ile Thr Gln Lys Lys Phe Leu Cys Asp Arg Thr Leu Trp Arg Ile Pro Phe Ser Ser Asn Phe Met Ser Met Gly Ala Leu Thr Asp Leu Gly Gln Asn Leu Leu Tyr Ala Asn 905

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Ser Ala His Ala Leu Asp Met Thr Phe Glu Val Asp Pro Met Asp Glu 920 Pro Thr Leu Leu Tyr Val Leu Phe Glu Val Phe Asp Val Val Arg Val 930 935 His Gln Pro His Arg Gly Val Ile Glu Thr Val Tyr Leu Arg Thr Pro 950 955 Phe Ser Ala Gly Asn Ala Thr Thr (2) INFORMATION FOR SEQ ID NO: 3: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2858 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (ix) FEATURE (A) NAME/KEY: misc_feature
(B) LOCATION: 951, 952 (D) OTHER INFORMATION: /note="Xaa can be either Gln, His, or Thr" (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3: ATG GCT ACC CCT TCG ATG ATG CCG CAG TGG TCT TAC ATG CAC ATC TCG 48 Met Ala Thr Pro Ser Met Met Pro Gln Trp Ser Tyr Met His Ile Ser 1 GGC CAG GAC GCC TCG GAG TAC CTG AGC CCC GGG CTG GTG CAG TTT GCC 96 Gly Gln Asp Ala Ser Glu Tyr Leu Ser Pro Gly Leu Val Gln Phe Ala CGC GCC ACC GAG ACG TAC TTC AGC CTG AAT AAC AAG TTT AGA AAC CCC 144 Arg Ala Thr Glu Thr Tyr Phe Ser Leu Asn Asn Lys Phe Arg Asn Pro ACG GTG GCG CCT ACG CAC GAC GTG ACC ACA GAC CGG TCC CAG CGT TTG 192 Thr Val Ala Pro Thr His Asp Val Thr Thr Asp Arg Ser Gln Arg Leu ACG CTG CGG TTC ATC CCT GTG GAC CGT GAG GAT ACT GCG TAC TCG TAC 240 Thr Leu Arg Phe Ile Pro Val Asp Arg Glu Asp Thr Ala Tyr Ser Tyr 70 288 AAG GCG CGG TTC ACC CTA GCT GTG GGT GAT AAC CGT GTG CTG GAC ATG Lys Ala Arg Phe Thr Leu Ala Val Gly Asp Asn Arg Val Leu Asp Met GCT TCC ACG TAC TTT GAC ATC CGC GGC GTG CTG GAC AGG GGC CCT ACT 336 Ala Ser Thr Tyr Phe Asp Ile Arg Gly Val Leu Asp Arg Gly Pro Thr 100 TTT AAG CCC TAC TCT GGC ACT GCC TAC AAC GCC CTG GCT CCC AAG GGT 384 Phe Lys Pro Tyr Ser Gly Thr Ala Tyr Asn Ala Leu Ala Pro Lys Gly 120 GCC CCA AAT CCT TGC GAA TGG GAT GAA GCT GCT ACT GCT CTT GAA ATA 432 Ala Pro Asn Pro Cys Glu Trp Asp Glu Ala Ala Thr Ala Leu Glu Ile

			GAT Asp						480
			CAC His						528
			GGT Gly						576
			ACA Thr						624
			GAA Glu 215						672
 			AAA Lys						720
			CAA Gln						768
			GAA Glu		Phe				816
			AAC Asn					AGT Ser	. 864
			ACC Thr 295						912
			TCA Ser						960
			ATT Ile						1008
 	 	 	GGT Gly	 		 		 GCA Ala	1056
								CTT Leu	1104
			GAT Asp 375						1152
Met			Val					ATT Ile 400	1200

			CAT His														1248
CTG Leu	GGA Gly	GGT Gly	GTG Val 420	ATT Ile	AAT Asn	ACA Thr	GAG Glu	ACT Thr 425	CTT Leu	ACC Thr	AAG Lys	GTA Val	AAA Lys 430	CCT Pro	AAA Lys		1296
			GAA Glu														1344
			ATA Ile														1392
			CTG Leu														1440
TTG Leu	CCC Pro	GAC Asp	AAG Lys	CTA Leu 485	AAG Lys	TAC Tyr	AGT Ser	CCT Pro	TCC Ser 490	AAC Asn	GTA Val	AAA Lys	ATT Ile	TCT Ser 495	GAT Asp		1488
AAC Asn	CCA Pro	AAC Asn	ACC Thr 500	TAC Tyr	GAC Asp	TAC Tyr	ATG Met	AAC Asn 505	AAG Lys	CGA Arg	GTG Val	GTG Val	GCT Ala 510	CCC Pro	GGG Gly		1536
			TGC Cys														1584
Met	GAC Asp 530	AAC Asn	GTC Val	AAC Asn	CCA Pro	TTT Phe 535	AAC Asn	CAC His	CAC His	CGC Arg	AAT Asn 540	GCT Ala	GGC Gly	CTG Leu	CGC Arg	•	1632
	Arg		ATG Met														1680
			CAG Gln		Phe												1728
GGC Gly	TCA Ser	TAC Tyr	ACC Thr 580	TAC Tyr	GAG Glu	TGG Trp	AAC Asn	TTC Phe 585	AGG Arg	AAG Lys	GAT Asp	GTT Val	AAC Asn 590	Met	GTT Val		1776
			TCC					Leu					Ala				1824
		Asp	AGC Ser				Tyr					Pro					1872
	Thr		TCC Ser			Glu					Asn				GAC Asp 640		1920
					Tyr					Asn					ATA Ile		1968

			-													
CCC Pro	GCC Ala	AAC Asn	GCT Ala 660	ACC Thr	AAC Asn	GTG Val	CCC Pro	ATA Ile 665	TCC Ser	ATC Ile	CCC Pro	TCC Ser	CGC Arg 670	AAC Asn	TGG Trp	2016
GCG Ala	GCT Ala	TTC Phe 675	CGC Arg	GGC Gly	TGG Trp	GCC Ala	TTC Phe 680	ACG Thr	CGC Arg	CTT Leu	AAG Lys	ACT Thr 685	AAG Lys	GAA Glu	ACC Thr	2064
	TCA Ser 690															2112
	CCC Pro															2160
	GCC Ala															2208
	CTT Leu															2256
	TAC Tyr															2304
	ATG Met 770															2352
	AGC Ser															2400
	AGC Ser															2448
	GGC Gly															2496
	CCC Pro		Met						_	_		_		_	_	2544
	CTT Leu 850															2592
	TGC Cys															2640
	ATG Met															2688
	GCC Ala			Leu					Glu							2736

									70								
CCC Pro	ACC Thr	CTT Leu 915	CTT Leu	TAT Tyr	GTT Val	TTG Leu	TTT Phe 920	GAA Glu	GTC Val	TTT Phe	GAC Asp	GTG Val 925	GTC Val	CGT Arg	GTG Val		2784
CAC His	CGG Arg 930	CCG Pro	CAC His	CGC Arg	GGC Gly	GTC Val 935	ATC Ile	GAA Glu	ACC Thr	GTG Val	TAC Tyr 940	CTG Leu	CGC Arg	ACG Thr	CCC Pro		2832
		GCC Ala						нн									2858
(2)	INFO	RMAT	ION	FOR	SEQ	ID N	iO:4:										
	(i)	(B	UENC) LE) TY)) TC	NGTH	l: 95 amir	2 am	ino id		is								
	(ii)	MOL	ECUL	E TY	PE:	prot	ein										
Thr _.		(E	A) NA B) LC	ME/F	ON:	951,	952			"Xaa	a cai	n be	eith	ner (Gln,	His,	or
	(xi)	SEÇ	QUENC	E DE	ESCR	PTIC	on: S	SEQ :	ID NO	0:4:	•						
Met 1	Ala	Thr	Pro	Ser 5	Met	Met	Pro	Gln	Trp 10	Ser	Tyr	Met	His	Ile 15	Ser		
Gly	Gln	Asp	Ala 20	Ser	Glu	Tyr	Leu	Ser 25	Pro	Gly	Leu	Val	Gln 30	Phe	Ala		
Arg	Ala	Thr 35	Glu	Thr	Tyr	Phe	Ser 40	Leu	Asn	Asn	Lys	Phe 45	Arg	Asn	Pro		
Thr	Val 50	Ala	Pro	Thr	His	Asp 55	Val	Thr	Thr	Asp	Arg 60	Ser	Gln	Arg	Leu		
Thr 65		Arg	Phe	Ile	Pro 70	Val	Asp	Arg	Glu	Asp 75	Thr	Ala	Tyr	Ser	Tyr 80		
Lys	Ala	Arg	Phe	Thr 85	Leu	Ala	Val	Gly	Asp 90	Asn	Arg	Val	Leu	Asp 95			-
Ala	Ser	Thr	Tyr 100	Phe	Asp	Ile	Arg	Gly 105		Leu	Asp	Arg	Gly 110	Pro	Thr		
Phe	Lys	Pro 115	Tyr	Ser	Gly	Thr	Ala 120		Asn	Ala	Leu	Ala 125		Lys	Gly		
Ala	Pro 130		Pro	Cys	Glu	Trp 135		Glu	Ala	Ala	Thr 140		Leu	Glu	Ile		
Asn 145		Glu	Glu	Glu	Asp 150		Asp	Asn	Glu	Asp 155		Val	Asp	Glu	Gln 160		
Ala	Glu	Gln	Gln	Lys 165		His	Val	Phe	Gly 170		Ala	Pro	Tyr	Ser 175	Gly		
Ile	Asn	Ile	Thr 180	_	Glu	Gly	Ile	Gln 185		Gly	Val	Glu	Gly 190		Thr	•	

Pro	Lys	Tyr 195	Ala	Asp	Lys	Thr	Phe 200	Gln	Pro	Glu	Pro	Gln 205	Ile	Gly	Glu
Ser	Gln 210	Trp	Tyr	Glu	Thr	Glu 215	Ile	Asn	His	Ala	Ala 220	Gly	Arg	Val	Leu
Lys 225	Lys	Thr	Thr	Pro	Met 230	Lys	Pro	Cys	Tyr	Gly 235	Ser	Tyr	Ala	Lys	Pro 240
Thr	Asn	Glu	Asn	Gly 245	Gly	Gln	Gly	Ile	Leu 250	Val	Lys	Gln	Gln	Asn 255	Gly
Lys	Leu	Glu	Ser 260	Gln	Val	Glu	Met	Gln 265	Phe	Phe	Ser	Thr	Thr 270	Glu	Ala
Thr	Ala	Gly 275	Asn	Gly	Asp	Asn	Leu 280	Thr	Pro	Lys	Val	Val 285	Leu	Tyr	Ser
Glu	Asp 290	Val	Asp	Ile	Glu	Thr 295	Pro	Asp	Thr	His	Ile 300	Ser	Tyr	Met	Pro
Thr 305	Ile	Lys	Glu	Gly	Asn 310	Ser	Arg	Glu	Leu	Met 315	Gly	Gln	Gln	Ser	Met 320
Pro	Asn	Arg	Pro	Asn 325	Tyr	Ile	Ala	Phe	Arg 330	Asp	Asn	Phe		Gly 335	Leu
Met	Tyr	Tyr	Asn 340	Ser	Thr	Gly	Asn	Met 345	Gly	Val	Leu	Ala	Gly 350	Gln	Ala
Ser	Gln	Leu 355	Asn	Ala	Val	Val	Asp 360	Leu	Gln	Asp	Arg	Asn 365	Thr	Glu	Leu
Ser	Tyr 370	Gln	Leu	Leu	Leu	Asp 375	Ser	Ile	Gly	Asp	Arg 380	Thr	Arg	Tyr	Phe
Ser 385	Met	Trp	Asn	Gln	Ala 390	Val	Asp	Ser	Tyr	Asp 395	Pro	Asp	Val	Arg	Ile 400
Ile	Glu	Asn	His	Gly 405	Thr	Glu	Asp	Glu	Leu 410	Pro	Asn	Tyr	Cys	Phe 415	Pro
Leu	Gly	Gly	Val 420	Ile	Asn	Thr	Glu	Thr 425	Leu	Thr	Lys	Val	Lys 430	Pro	Lys
Thr	Gly	Gln 435	Glu	Asn	Gly	Trp	Glu 440	Lys	Asp	Ala	Thr	Glu 445	Phe	Ser	Asp
Lys	Asn 450	Glu	Ile	Arg	Val	Gly 455	Asn	Asn	Phe	Ala	Met 460	Glu	Ile	Asn	Leu
Asn 465	Ala	Asn	Leu	Trp	Arg 470	Asn	Phe	Leu	Tyr	Ser 475	Asn	Ile	Ala	Leu	Tyr 480
Leu	Pro	Asp	Lys	Leu 485	Lys	Tyr	Ser	Pro	Ser 490	Asn	Val	Lys	Ile	Ser 495	Asp
Asn	Pro	Asn	Thr 500	Tyr	Asp	Tyr	Met	Asn 505	Lys	Arg	Val	Val	Ala 510	Pro	Gly
Leu	Val	Asp 515	Cys	Tyr	Ile	Asn	Leu 520	Gly	Ala	Arg	Trp	Ser 525	Leu	Asp	Tyr

Met	Asp 530	Asn	Val	Asn	Pro	Phe 535	Asn	His	His	Arg	Asn 540	Ala	Gly	Leu	Arg
Tyr 545	Arg	Ser	Met	Leu	Leu 550	Gly	Asn	Gly	Arg	Tyr 555	Val	Pro	Phe	His	Ile 560
Gln	Val	Pro	Gln	Lys 565	Phe	Phe	Ala	Ile	Lys 570	Asn	Leu	Leu	Leu	Leu 575	Pro
Gly	Ser	Tyr	Thr 580	Tyr	Glu	Trp	Asn	Phe 585	Arg	Lys	Asp	Val	Asn 590	Met	Val
Leu	Gln	Ser 595	Ser	Leu	Gly	Asn	Asp 600	Leu	Arg	Val	Asp	Gly 605	Ala	Ser	Ile
Lys	Phe 610	Asp	Ser	Ile	Cys	Leu 615	Tyr	Ala	Thr	Phe	Phe 620	Pro	Met	Ála	His
Asn 625	Thr	Ala	Ser	Thr	Leu 630	Glu	Ala	Met	Leu	Arg 635	Asn	Asp	Thr	Asn	Asp 640
Gln	Ser	Phe	Asn	Asp 645	Tyr	Leu	Ser	Ala	Ala 650	Asn	Met	Leu	Tyr	Pro 655	Ile
Pro	Ala	Asn	Ala 660	Thr	Asn	Val	Pro	Ile 665	Ser	Ile	Pro	Ser	Arg 670	Asn	Trp
Ala	Ala	Phe 675	Arg	Gly	Trp	Ala	Phe 680	Thr	Arg	Leu	Lys	Thr 685	Lys	Glu	Thr
Pro	Ser 690	Leu	Gly	Ser	Gly	Tyr 695	Asp	Pro	Tyr	Tyr	Thr 700		Ser	Gly	Ser
Ile 705	Pro	Tyr	Leu	Asp	Gly 710	Thr	Phe	Tyr	Leu	Asn 715	His	Thr	Phe	Lys	Lys 720
Val	Ala	Ile	Thr	Phe 725	Asp	Ser	Ser	Val	Ser 730	Trp	Pro	Gly	Asn	Asp 735	Arg
Leu	Leu	Thr	Pro .740	Asn	Glu	Phe	Glu	Ile 745	Lys	Arg	Ser	Val	Asp 750	Gly	Glu
Gly	Tyr	Asn 755	Val	Ala	Gln	Суѕ	Asn 760	Met	Thr	Lys	Asp	Trp 765	Phe	Leu	Val
Gln	Met 770	Leu	Ala	Asn	Tyr	Asn 775	Ile	Gly	Tyr	Gln	Gly 780	Phe	Tyr	Ile	Pro
Glu 785	Ser	Tyr	Lys	Asp	Arg 790	Met	Tyr	Ser	Phe	Phe 795	Arg	Asn	Phe	Gln	Pro 800
Met	Ser	Arg	Gln	Val 805	Val	Asp	Asp	Thr	Lys 810	Tyr	Lys	Asp	Tyr	Gln 815	Gln
Val	Gly	Ile	Leu 820	His	Gln	His	Asn	Asn 825	Ser	Gly	Phe	Val	Gly 830	Tyr	Leu
Ala	Pro	Thr 835	Met	Arg	Glu	Gly	Gln 840	Ala	Tyr	Pro	Ala	Asn 845	Phe	Pro	Tyr
Pro	Leu 850	Ile	Gly	Lys	Thr	Ala 855	Val	Asp	Ser	Ile	Thr 860	Gln	·Lys	Lys	Phe

Leu 865	Cys	Asp	Arg	Thr	Leu 870	Trp	Arg	Ile	Pro	Phe 875	Ser	Ser	Asn	Phe	Met 880	·
Ser	Met	Gly	Ala	Leu 885	Thr	Asp	Leu	Gly	Gln 890	Asn	Leu	Leu	Tyr	Ala 895	Asn	
Ser	Ala	His	Ala 900	Leu	Asp	Met	Thr	Phe 905	Glu	Val	Asp	Pro	Met 910	Asp	Glu	
Pro	Thr	Leu 915	Leu	Tyr	Val	Leu	Phe 920	Glu	Val	Phe	Asp	Val 925	Val	Arg	Val	
His	Arg 930	Pro	His	Arg	Gly	Val 935	Ile	Glu	Thr	Val	Tyr 940	Leu	Arg	Thr	Pro	
Phe 945	Ser	Ala	Gly	Asn	Ala 950	Xaa	Xaa				-					
(2)	INFO	RMAI	NOI	FOR	SEQ	ID N	10:5:	:								
	(i)	(<i>P</i> (<i>E</i>	QUENC A) LE B) TY C) SI C) TO	NGTI PE: RANI	i: 60 nucl	03 ba Leic ESS:	ase p acid doub	oairs i	3							
	(ii)	MOI	LECUI	E TY	PE:	DNA	(ger	nomic	=)							
	(xi)	SEC	QUENC	E DE	ESCR	PTIC	ON: S	SEQ 1	D. NO	0:5:						
	TGT Cys															48
	GAA Glu															96
	GCT Ala															144
	TTG Leu 50															192
	AAT Asn															240
	CCA Pro			-												288
	GCG Ala															336
	TAT Tyr		Ser					Thr					Gly			384

GTT Val	CTG Leu 130	Val	CCG Pro	GAT Asp	GAA Glu	AAA Lys 135	GGG Gly	GTG Val	CCT Pro	CTT Leu	CCA Pro 140	AAG Lys	GTT Val	GAC Asp	TTG Leu	432
CAA Gln 145	TTC Phe	TTC Phe	TCA Ser	AAT Asn	ACT Thr 150	ACC Thr	TCT Ser	TTG Leu	AAC Asn	GAC Asp 155	CGG Arg	CAA Gln	GGC Gly	AAT Asn	GCT Ala 160	480
ACT Thr	AAA Lys	CCA Pro	AAA Lys	GTG Val 165	GTT Val	TTG Leu	TAC Tyr	AGT Ser	GAA Glu 170	GAT Asp	GTA Val	AAT Asn	ATG Met	GAA Glu 175	ACC Thr	528
CCA Pro	GAC Asp	ACA Thr	CAT His 180	CTG Leu	TCT Ser	TAC Tyr	AAA Lys	CCT Pro 185	GGA Gly	AAA Lys	GGT Gly	GAT Asp	GAA Glu 190	AAT Asn	TCT Ser	576
AAA Lys					CAA Gln											603
(2)	INFO	RMA	CION	FOR	SEQ	ID N	10:6	:				•				
	(i)	(<i>I</i>	A) LE 3) TY	ENGTI (PE:	ARAC 1: 20 amir OGY:)l an no ac	nino cid		is							
	·				(PE: ESCRI			SEQ I	ED NO	0:6:						
Ser 1	Cys	Glu	Trp	Glu 5	Gln	Thr	Glu	Asp	Ser 10	Gly	Arg	Ala	Val	Ala 15	Glu	•
Asp	Glu	Glu	Glu 20	Glu	Asp	Glu	Asp	Glu 25	Glu	Glu	Glu	Glu	Glu 30	Glu	Gln	-
Asn	Ala	Arg 35	Asp	Gln	Ala	Thr	Lys 40	Lys	Thr	His	Val	Tyr 45	Ala	Gln	Ala	
Pro	Leu 50	Ser	Gly	Glu	Thr	Ile 55	Thr	Lys	Ser	Gly	Leu 60	Gln	Ile	Gly	Ser	
Asp 65	Asn	Ala	Glu	Thr	Gln 70	Ala	Lys	Pro	Val	Tyr 75	Ala	Asp	Pro	Ser	Tyr 80	
Gln	Pro	Glu	Pro	Gln 85	Ile	Gly	Glu	Ser	Gln 90		Asn	Glu	Ala	Asp 95	Ala	
Asn	Ala	Ala	Gly 100		Arg	Val	Leu	Lys 105	Lys	Thr	Thr	Pro	Met 110		Pro	
Суѕ	Tyr	Gly 115		Tyr	Ala	Arg	Pro 120		Asn	Pro	Phe	Gly 125		Gln	Ser	
Val	Leu 130		Pro	Asp	Glu	Lys 135		Val	Pro	Leu	Pro 140		Val	Asp	Leu	
Gln 145		Phe	Ser	Asn	Thr 150		Ser	Leu	Asn	Asp 155		Gln	Gly	Asn	Ala 160	
Thr	Lys	Pro	Lys	Val 165		Leu	Туг	Ser	Glu 170		Val	Asn	Met	Glu 175	Thr	

Pro Asp Thr His Leu Ser Tyr Lys Pro Gly Lys Gly Asp Glu Asn Ser 185

Lys Ala Met Leu Gly Gln Gln Ser Met 195 200

- (2) INFORMATION FOR SEQ ID NO:7:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 567 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double

 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

	(xi)	SEÇ	DUENC	E DE	ESCR	PTIC	on: S	SEQ 1	D NC):/:					
	TGC Cys														48
	GAG Glu														96
	AAA Lys														144
	AAG Lys 50														192
	GAT Asp														240
	GAA Glu														288
	CCA Pro														336
	GGA Gly													GAA Glu	384
	CAA Gln 130														432
	GGT Gly														480
	ATA Ile													AAG Lys	528
GAA	GGT	AAC	TCA	CGA	GAA	CTA	ATG	GGC	CAA	CAA	TCT	ATG			567

Glu Gly Asn Ser Arg Glu Leu Met Gly Gln Gln Ser Met 180 185

- (2) INFORMATION FOR SEQ ID NO:8:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 189 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Pro Cys Glu Trp Asp Glu Ala Ala Thr Ala Leu Glu Ile Asn Leu Glu 1 5 10 15.

Gln Lys Thr His Val Phe Gly Gln Ala Pro Tyr Ser Gly Ile Asn Ile $35 \hspace{1cm} 40 \hspace{1cm} 45$

Thr Lys Glu Gly Ile Gln Ile Gly Val Glu Gly Gln Thr Pro Lys Tyr
50 55 60

Ala Asp Lys Thr Phe Gln Pro Glu Pro Gln Ile Gly Glu Ser Gln Trp 65 70 75 80

Tyr Glu Thr Glu Ile Asn His Ala Ala Gly Arg Val Leu Lys Lys Thr 85 90 95

Thr Pro Met Lys Pro Cys Tyr Gly Ser Tyr Ala Lys Pro Thr Asn Glu 100 105 110

Asn Gly Gly Gln Gly Ile Leu Val Lys Gln Gln Asn Gly Lys Leu Glu 115 120 125

Ser Gln Val Glu Met Gln Phe Phe Ser Thr Thr Glu Ala Thr Ala Gly 130 135 140

Asn Gly Asp Asn Leu Thr Pro Lys Val Val Leu Tyr Ser Glu Asp Val 145 150 155 160

Asp Ile Glu Thr Pro Asp Thr His Ile Ser Tyr Met Pro Thr Ile Lys 165 170 175

Glu Gly Asn Ser Arg Glu Leu Met Gly Gln Gln Ser Met 180 185

- (2) INFORMATION FOR SEQ ID NO:9:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 153 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

ACC GAA GAT AGC GGC CGG GCA GTT GCC GAG GAT GAA GAA GAG GAA GAT

Thr 1	Glu	Asp	Ser	Gly 5	Arg	Ala	Val	Ala	Glu 10	Asp	Glu	Glu	Glu	Glu 15	Asp	
				GAG Glu												96
				CAT His												144
	ACA Thr 50															153
(2)	INFO	RMA	NOIT	FOR	SEQ	ID N	10:10):						-		
	(i)	(I	A) LE B) TY	CE CH ENGTH (PE: OPOLO	l: 51 amir	ami	no a		5		٠	,				
	(ii)	MOI	LECUI	LE TY	PE:	pept	ide									•
	(xi)	SE	QUENC	CE DE	SCR	PTIC	ON: S	SEQ 1	D NO	0:10:	:					
Thr 1	Glu	Asp	Ser	Gly 5	Arg	Ala	Val	Ala	Glu 10	Asp	Glu	Glu	Glu	Glu 15	Asp	
Glu	Asp	Glu	Glu 20	Glu	Glu	Glu	Glu	Glu 25	Gln	Asn	Ala	Arg	Asp 30	Gln	Ala	
Thr	Lys	Lys 35	Thr	His	Val	Tyr	Ala 40	Gln	Ala	Pro	Leu	Ser 45	Gly	Glu	Thr	
Ile	Thr 50	Lys										,				
(2)	INFO	ORMA!	rion	FOR	SEQ	ID I	10:13	l:		•						
	(i)	() (1	A) L1 B) T' C) S'	CE CH ENGTH YPE: TRANI OPOLO	i: 1: nuc. DEDNI	35 ba leic ESS:	ase p acid doub	pair:	5							
	(ii)	MO:	LECU	LE TY	PE:	DNA	(ge	nomi	c)	•						
	(xi	SE	QUEN	CE DI	ESCR	[PTI	วท: :	SEQ :	ID N	0:11	:					
				CTT Leu 5											Asn	48
				GAC Asp												96
			Pro	TAT Tyr												135

- (2) INFORMATION FOR SEQ ID NO:12:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 45 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Ala Ala Thr Ala Leu Glu Ile Asn Leu Glu Glu Glu Asp Asp Asn 1 5 10 15

Glu Asp Glu Val Asp Glu Gln Ala Glu Gln Gln Lys Thr His Val Phe 20 25 30

Gly Gln Ala Pro Tyr Ser Gly Ile Asn Ile Thr Lys Glu 35 40 45

- (2) INFORMATION FOR SEQ ID NO:13:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 33 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

TCA GAC AAT GCA GAA ACA CAA GCT AAA CCT GTA Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val 1 5 10

- (2) INFORMATION FOR SEQ ID NO:14:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 11 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val 1 5 10

- (2) INFORMATION FOR SEQ ID NO:15:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

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GTC GAA GGT CAA ACA CCT AAA 21 Val Glu Gly Gln Thr Pro Lys 1. (2) INFORMATION FOR SEQ ID NO:16: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 7 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: peptide (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16: Val Glu Gly Gln Thr Pro Lys (2) INFORMATION FOR SEQ ID NO:17: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 24 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17: AAC GAA GCT GAT GCT AAT GCG GCA 24 Asn Glu Ala Asp Ala Asn Ala Ala 1 5 (2) INFORMATION FOR SEQ ID NO:18: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 8 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: peptide (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18: Asn Glu Ala Asp Ala Asn Ala Ala (2) INFORMATION FOR SEQ ID NO:19: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 24 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19: TAC GAA ACT GAA ATT AAT CAT GCA

Tyr Glu Thr Glu Ile Asn His Ala

- (2) INFORMATION FOR SEQ ID NO:20:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Tyr Glu Thr Glu Ile Asn His Ala

- (2) INFORMATION FOR SEQ ID NO:21:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 42 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

TCC GTT CTG GTT CCG GAT GAA AAA GGG GTG CCT CTT CCA AAG
Ser Val Leu Val Pro Asp Glu Lys Gly Val Pro Leu Pro Lys
1 5 10

42

- (2) INFORMATION FOR SEQ ID NO:22:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 14 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Ser Val Leu Val Pro Asp Glu Lys Gly Val Pro Leu Pro Lys
1 5 10

- (2) INFORMATION FOR SEQ ID NO:23:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 42 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

GGC ATT CTT GTA AAG CAA CAA AAT GGA AAG CTA GAA AGT CAA
Gly Ile Leu Val Lys Gln Gln Asn Gly Lys Leu Glu Ser Gln
1 10

- (2) INFORMATION FOR SEQ ID NO:24:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 14 amino acids

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AAA

Lys

1

Lys

81

- (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: peptide (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24: Gly Ile Leu Val Lys Gln Gln Asn Gly Lys Leu Glu Ser Gln (2) INFORMATION FOR SEQ ID NO:25: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 51 base pairs (B) TYPE: nucleic acid(C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25: TCA AAT ACT ACC TCT TTG AAC GAC CGG CAA GGC AAT GCT ACT AAA CCA Ser Asn Thr Thr Ser Leu Asn Asp Arg Gln Gly Asn Ala Thr Lys Pro 5 51 (2) INFORMATION FOR SEQ ID NO:26: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 17 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: peptide (xi) SEQUENCE DESCRIPTION: SEO ID NO:26: Ser Asn Thr Thr Ser Leu Asn Asp Arg Gln Gly Asn Ala Thr Lys Pro
 - - (i) SEQUENCE CHARACTERISTICS:

(2) INFORMATION FOR SEQ ID NO:27:

- (A) LENGTH: 48 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

TCA ACT ACT GAG GCG ACC GCA GGC AAT GGT GAT AAC TTG ACT CCT AAA 48 Ser Thr Thr Glu Ala Thr Ala Gly Asn Gly Asp Asn Leu Thr Pro Lys 10

- (2) INFORMATION FOR SEQ ID NO:28:
 - (i) SEQUENCE CHARACTERISTICS:

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- (A) LENGTH: 16 amino acids
 - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Ser Thr Thr Glu Ala Thr Ala Gly Asn Gly Asp Asn Leu Thr Pro Lys 10

- (2) INFORMATION FOR SEQ ID NO:29:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

TTG TAC AGT GAA GAT GTA AAT ATG Leu Tyr Ser Glu Asp Val Asn Met

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- (2) INFORMATION FOR SEQ ID NO:30:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid(D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Leu Tyr Ser Glu Asp Val Asn Met

- (2) INFORMATION FOR SEQ ID NO:31:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

TTG TAC AGT GAA GAT GTA GAT ATA Leu Tyr Ser Glu Asp Val Asp Ile

- (2) INFORMATION FOR SEQ ID NO:32:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 (D) TOPOLOGY: linear

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- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Leu Tyr Ser Glu Asp Val Asp Ile

- (2) INFORMATION FOR SEQ ID NO:33:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 36 base pairs

 - (B) TYPE: nucleic acid(C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

GGA AAA GGT GAT GAA AAT TCT AAA GCT ATG TTG GGT Gly Lys Gly Asp Glu Asn Ser Lys Ala Met Leu Gly 36

- (2) INFORMATION FOR SEQ ID NO:34:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 12 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:
- Gly Lys Gly Asp Glu Asn Ser Lys Ala Met Leu Gly
- (2) INFORMATION FOR SEQ ID NO:35:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 36 base pairs (B) TYPE: nucleic acid

 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

ACT ATT AAG GAA GGT AAC TCA CGA GAA CTA ATG GGC Thr Ile Lys Glu Gly Asn Ser Arg Glu Leu Met Gly

- (2) INFORMATION FOR SEQ ID NO:36:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 12 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Thr Ile Lys Glu Gly Asn Ser Arg Glu Leu Met Gly $1 \hspace{1cm} 5 \hspace{1cm} 10$

- (2) INFORMATION FOR SEQ ID NO: 37:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 165 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

AAT TAT TGT TTT CCT CTT GGG GGT ATT GGG GTA ACT GAC ACC TAT CAA
Asn Tyr Cys Phe Pro Leu Gly Gly Ile Gly Val Thr Asp Thr Tyr Gln
1 5 10 15

GCT ATT AAG GCT AAT GGC AAT GGC TCA GGC GAT AAT GGA GAT ACT ACA
Ala Ile Lys Ala Asn Gly Asn Gly Ser Gly Asp Asn Gly Asp Thr Thr
20 25 30

TGG ACA AAA GAT GAA ACT TTT GCA ACA CGT AAT GAA ATA GGA GTG GGT
Trp Thr Lys Asp Glu Thr Phe Ala Thr Arg Asn Glu Ile Gly Val Gly
35 40 45

AAC AAC TTT GCC ATG GAA ATT Asn Asn Phe Ala Met Glu Ile 50 55 165

- (2) INFORMATION FOR SEQ ID NO:38:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 55 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

Asn Tyr Cys Phe Pro Leu Gly Gly Ile Gly Val Thr Asp Thr Tyr Gln
1 5 10 15

Ala Ile Lys Ala Asn Gly Asn Gly Ser Gly Asp Asn Gly Asp Thr Thr 20 25 30

Trp Thr Lys Asp Glu Thr Phe Ala Thr Arg Asn Glu Ile Gly Val Gly 35 40 45

Asn Asn Phe Ala Met Glu Ile 50 55

- (2) INFORMATION FOR SEQ ID NO:39:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 153 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)

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	(xi) SEQUENCE DESCRIPTION: S	EQ ID NO	:39:		
AAT Asn 1	T TAC TGC TTT CCA CTG GGA GGT n Tyr Cys Phe Pro Leu Gly Gly 1 5	GTG ATT A Val Ile A 10	AAT ACA GAG Asn Thr Glu	ACT CTT A Thr Leu T 15	CC 48 hr
	G GTA AAA CCT AAA ACA GGT CAG s Val Lys Pro Lys Thr Gly Gln 20				
	A GAA TTT TCA GAT AAA AAT GAA r Glu Phe Ser Asp Lys Asn Glu 35 40				
	G GAA ATC t Glu Ile 50				153
(2)) INFORMATION FOR SEQ ID NO:40	:			
	(i) SEQUENCE CHARACTERISTIC (A) LENGTH: 51 amino a (B) TYPE: amino acid (D) TOPOLOGY: linear			·	
	(ii) MOLECULE TYPE: peptide				
	(xi) SEQUENCE DESCRIPTION: S	EQ ID NO	:40:		
Asn 1	n Tyr Cys Phe Pro Leu Gly Gly 1 5	Val Ile A 10	Asn Thr Glu	Thr Leu Ti	hr
	s Val Lys Pro Lys Thr Gly Gln 20	25		30	
Thr	r Glu Phe Ser Asp Lys Asn Glu 35 40	Ile Arg '	Val Gly Asn 45	Asn Phe A	la
Met	t Glu Ile 50				
(2)) INFORMATION FOR SEQ ID NO:41	:			
	(i) SEQUENCE CHARACTERISTIC (A) LENGTH: 54 base pa (B) TYPE: nucleic acid (C) STRANDEDNESS: doub (D) TOPOLOGY: linear	irs			
	(ii) MOLECULE TYPE: DNA (gen	omic)			
	(xi) SEQUENCE DESCRIPTION: S	EQ ID NO	:41:		
GTA Val 1	A ACT GAC ACC TAT CAA GCT ATT 1 Thr Asp Thr Tyr Gln Ala Ile 1 5	AAG GCT A Lys Ala A 10	AAT GGC AAT Asn Gly Asn	GGC TCA G Gly Ser G 15	GC 48 ly
	T AAT p Asn				54
(2)) INFORMATION FOR SEQ ID NO:42	:			

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 amino acids

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- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

Val Thr Asp Thr Tyr Gln Ala Ile Lys Ala Asn Gly Asn Gly Ser Gly

Asp Asn

- (2) INFORMATION FOR SEQ ID NO:43:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 87 base pairs
 - (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

AAT ACA GAG ACT CTT ACC AAG GTA AAA CCT AAA ACA GGT CAG GAA AAT Asn Thr Glu Thr Leu Thr Lys Val Lys Pro Lys Thr Gly Gln Glu Asn

GGA TGG GAA AAA GAT GCT ACA GAA TTT TCA GAT AAA AAT Gly Trp Glu Lys Asp Ala Thr Glu Phe Ser Asp Lys Asn 20

- (2) INFORMATION FOR SEQ ID NO:44:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

Asn Thr Glu Thr Leu Thr Lys Val Lys Pro Lys Thr Gly Gln Glu Asn

- Gly Trp Glu Lys Asp Ala Thr Glu Phe Ser Asp Lys Asn 20
 - (2) INFORMATION FOR SEQ ID NO:45:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 base pairs(B) TYPE: nucleic acid

 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

ACT TTT GCA ACA CGT AAT GAA Thr Phe Ala Thr Arg Asn Glu 21

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(2) INFORMATION FOR SEQ ID NO:46:
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- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

Thr Phe Ala Thr Arg Asn Glu

- (2) INFORMATION FOR SEQ ID NO: 47:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

ACA GAA TTT TCA GAT AAA AAT GAA Thr Glu Phe Ser Asp Lys Asn Glu 24

- (2) INFORMATION FOR SEQ ID NO:48:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

Thr Glu Phe Ser Asp Lys Asn Glu

- (2) INFORMATION FOR SEQ ID NO:49:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: other nucleic acid
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:

GAC TAC AAA GAC GAC GAC GAC AAA Asp Tyr Lys Asp Asp Asp Lys

- (2) INFORMATION FOR SEQ ID NO:50:
 - (i) SEQUENCE CHARACTERISTICS:

		(B) TY	PE:	: 8 amin GY:	o ac	id	ids					-	
	(ii)	MOL	ECUL	E TY	PE:	pept	ide							
	(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ I	D NO	:50:				
Asp 1	Tyr	Lys	Asp	Asp 5	Asp	Asp	Lys							
(2)	INFO	RMAT	ION	FOR	SEQ	ID N	0:51	. :						
	(i)	(A (B	L) LE () TY () ST	NGTH PE: RANE	IARAC 1: 29 nucl EDNE	07 b eic SS:	ase acid doub	pair l	·s					
	(ii)	MOL	ECUL	E TY	PE:	DNA	(gen	omic	:)					
	(xi)	SEÇ	UENC	E DE	SCRI	PTIC	N: S	EQ I	D NC	:51:				
ATG	GCT Ala 1				ATG Met 5						 			48
	CAG Gln													96
	GCC Ala													144
	GTG Val													192
	CTG Leu 65													240
	GCG Ala													28 8
	TCC Ser													336
	AAG Lys			Ser										384
	CCT Pro													432
	GCC Ala 145													480

		CGA Arg 165						528
		TCT Ser						576
		GCA Ala						624
		GAA Glu						672
		GCA Ala						720
		GGA Gly 245						768
		GTT Val						816
		TTC Phe						864
		CCA Pro						912
		ACA Thr						960
		ATG Met 325						1008
		AGG Arg						1056
		GGT Gly						1104
		CAA Gln						1152
		GGT Gly					AAT Asn	1200
		TAT Tyr 405						1248

ACT Thr								1296
GTA Val						 		1344
GAT Asp								1392
AAT Asn 465								1440
GCC Ala								1488
CCA Pro								1536
CCC Pro								1584
GTA Val								1632
GAC Asp 545								1680
CGC Arg								1728
GTG Val	Gln							1776
TCA Ser								1824
CAG Gln								1872
TTT Phe 625								1920
ACG Thr								1968
TCC Ser								2016

					AAC Asn											2064
					TGG Trp						-					2112
					GGC Gly											2160
					GGA Gly 725											2208
GTG Val	GCC Ala	ATT Ile	ACC Thr	TTT Phe 740	GAC Asp	TCT Ser	TCT Ser	GTT Val	AGC Ser 745	TGG Trp	CCG Pro	GGC Gly	AAC Asn	GAC Asp 750	CGC Arg	2256
					GAG Glu											2304
					CAG Gln											2352
					TAC Tyr											2400
					CGC Arg 805											2448
					GTT Val											2496
					CAG Gln											2544
					GAG Glu											2592
					ACC Thr											2640
					CTT Leu 885											2688
					ACA Thr											2736
					GAC Asp											2784

									12							
CCC Pro	ACC Thr	CTT Leu 930	CTT Leu	TAT Tyr	GTT Val	TTG Leu	TTT Phe 935	GAA Glu	GTC Val	TTT Phe	GAC Asp	GTG Val 940	GTC Val	CGT Arg	GTG Val	2832
CAC His	CAG Gln 945	CCG Pro	CAC His	CGC Arg	GGC Gly	GTC Val 950	ATC Ile	GAG Glu	ACC Thr	GTG Val	TAC Tyr 955	CTG Leu	CGC Arg	ACG Thr	CCC Pro	2880
					GCC Ala 965			TAA								2,907
(2)	INFO	ORMAT	NOI	FOR	SEQ	ID N	10:52	2:								
	(i)	(F	A) LE	NGT	iarac i: 96	57 am	nino		ls							

- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:
- Ala Thr Pro Ser Met Met Pro Gln Trp Ser Tyr Met His Ile Ser Gly
 1 5 10 15
- Gln Asp Ala Ser Glu Tyr Leu Ser Pro Gly Leu Val Gln Phe Ala Arg 20 25 30
- Ala Thr Glu Thr Tyr Phe Ser Leu Asn Asn Lys Phe Arg Asn Pro Thr 35 40 45
- Val Ala Pro Thr His Asp Val Thr Thr Asp Arg Ser Gln Arg Leu Thr 50 60
- Leu Arg Phe Ile Pro Val Asp Arg Glu Asp Thr Ala Tyr Ser Tyr Lys
 65 70 75 80
- Ala Arg Phe Thr Leu Ala Val Gly Asp Asn Arg Val Leu Asp Met Ala 85 90 95
- Ser Thr Tyr Phe Asp Ile Arg Gly Val Leu Asp Arg Gly Pro Thr Phe 100 105 110
- Lys Pro Tyr Ser Gly Thr Ala Tyr Asn Ala Leu Ala Pro Lys Gly Ala 115 120 125
- Pro Asn Ser Cys Glu Trp Glu Gln Thr Glu Asp Ser Gly Arg Ala Val 130 135 140
- Glu Gln Asn Ala Arg Asp Gln Ala Thr Lys Lys Thr His Val Tyr Ala 165 170 175
- Gln Ala Pro Leu Ser Gly Glu Thr Ile Thr Lys Ser Gly Leu Gln Ile 180 185 190
- Gly Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val Tyr Ala Asp Pro 195 200 205
- Ser Tyr Gln Pro Glu Pro Gln Ile Gly Glu Ser Gln Trp Asn Glu Ala 210 215 220

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Asp Ala Asn Ala Ala Gly Gly Arg Val Leu Lys Lys Thr Thr Pro Met 230 Lys Pro Cys Tyr Gly Ser Tyr Ala Arg Pro Thr Asn Pro Phe Gly Gly Gln Ser Val Leu Val Pro Asp Glu Lys Gly Val Pro Leu Pro Lys Val Asp Leu Gln Phe Phe Ser Asn Thr Thr Ser Leu Asn Asp Arg Gln Gly 280 Asn Ala Thr Lys Pro Lys Val Val Leu Tyr Ser Glu Asp Val Asn Met Glu Thr Pro Asp Thr His Leu Ser Tyr Lys Pro Gly Lys Gly Asp Glu Asn Ser Lys Ala Met Leu Gly Gln Gln Ser Met Pro Asn Arg Pro Asn 330 Tyr Ile Ala Phe Arg Asp Asn Phe Ile Gly Leu Met Tyr Tyr Asn Ser . 345 Thr Gly Asn Met Gly Val Leu Ala Gly Gln Ala Ser Gln Leu Asn Ala Val Val Asp Leu Gln Asp Arg Asn Thr Glu Leu Ser Tyr Gln Leu Leu Leu Asp Ser Ile Gly Asp Arg Thr Arg Tyr Phe Ser Met Trp Asn Gln Ala Val Asp Ser Tyr Asp Pro Asp Val Arg Ile Ile Glu Asn His Gly Thr Glu Asp Glu Leu Pro Asn Tyr Cys Phe Pro Leu Gly Gly Ile Gly Val Thr Asp Thr Tyr Gln Ala Ile Lys Ala Asn Gly Asn Gly Ser Gly Asp Asn Gly Asp Thr Thr Trp Thr Lys Asp Glu Thr Phe Ala Thr Arg Asn Glu Ile Gly Val Gly Asn Asn Phe Ala Met Glu Ile Asn Leu Asn Ala Asn Leu Trp Arg Asn Phe Leu Tyr Ser Asn Ile Ala Leu Tyr Leu Pro Asp Lys Leu Lys Tyr Asn Pro Thr Asn Val Glu Ile Ser Asp Asn Pro Asn Thr Tyr Asp Tyr Met Asn Lys Arg Val Val Ala Pro Gly Leu Val Asp Cys Tyr Ile Asn Leu Gly Ala Arg Trp Ser Leu Asp Tyr Met Asp Asn Val Asn Pro Phe Asn His His Arg Asn Ala Gly Leu Arg Tyr

Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile Gln Val Pro Gln Lys Phe Phe Ala Ile Lys Asn Leu Leu Leu Pro Gly Ser Tyr Thr Tyr Glu Trp Asn Phe Arg Lys Asp Val Asn Met Val Leu Gln Ser Ser Leu Gly Asn Asp Leu Arg Val Asp Gly Ala Ser Ile Lys 615 Phe Asp Ser Ile Cys Leu Tyr Ala Thr Phe Phe Pro Met Ala His Asn Thr Ala Ser Thr Leu Glu Ala Met Leu Arg Asn Asp Thr Asn Asp Gln Ser Phe Asn Asp Tyr Leu Ser Ala Ala Asn Met Leu Tyr Pro Ile Pro 660 Ala Asn Ala Thr Asn Val Pro Ile Ser Ile Pro Ser Arg Asn Trp Ala Ala Phe Arg Gly Trp Ala Phe Thr Arg Leu Lys Thr Lys Glu Thr Pro 695 Ser Leu Gly Ser Gly Tyr Asp Pro Tyr Tyr Thr Tyr Ser Gly Ser Ile Pro Tyr Leu Asp Gly Thr Phe Tyr Leu Asn His Thr Phe Lys Lys Val Ala Ile Thr Phe Asp Ser Ser Val Ser Trp Pro Gly Asn Asp Arg Leu Leu Thr Pro Asn Glu Phe Glu Ile Lys Arg Ser Val Asp Gly Glu Gly 760 Tyr Asn Val Ala Gln Cys Asn Met Thr Lys Asp Trp Phe Leu Val Gln Met Leu Ala Asn Tyr Asn Ile Gly Tyr Gln Gly Phe Tyr Ile Pro Glu Ser Tyr Lys Asp Arg Met Tyr Ser Phe Phe Arg Asn Phe Gln Pro Met 810 Ser Arg Gln Val Val Asp Asp Thr Lys Tyr Lys Glu Tyr Gln Gln Val Gly Ile Leu His Gln His Asn Asn Ser Gly Phe Val Gly Tyr Leu Ala Pro Thr Met Arg Glu Gly Gln Ala Tyr Pro Ala Asn Val Pro Tyr Pro 855 Leu Ile Gly Lys Thr Ala Val Asp Ser Ile Thr Gln Lys Lys Phe Leu 865 Cys Asp Arg Thr Leu Trp Arg Ile Pro Phe Ser Ser Asn Phe Met Ser 890

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Met	Gly	Ala	Leu 900	Thr	Asp	Leu	Gly	Gln 905	Asn	Leu	Leu	Tyr	Ala 910	Asn	Ser	
Ala	His	Ala 915	Leu	Asp	Met	Thr	Phe 920	Glu	Val	Asp	Pro	Met 925	Asp	Glu	Pro	
Thr	Leu 930	Leu	Tyr	Val	Leu	Phe 935	Glu	Val	Phe	Asp	Val 940	Val	Arg	Val	His	
Gln 945	Pro	His	Arg	Gly	Val 950	Ile	Glu	Thr	Val	Tyr 955	Leu	Arg	Thr	Pro	Phe 960	
Ser	Ala	Gly	Asn	Ala 965	Thr	Thr								•	-	
(2)	INFO	ORMAI	CION	FOR	SEQ	ID N	10:53	3:								
	(i)	(F (C	A) LE B) T'(C) ST	ENGTI PE: PRANI	HARAC H: 28 nucl DEDNE DGY:	358 b Leic ESS:	ase acio douk	pai: 1	rs							
	(ii)	MOI	LECUI	LE T	YPE:	DNA	(ger	nomic	2)							
	(xi)	SEC	QUENC	CE DI	ESCRI	PTIC	on: s	SEQ 1	D NO	5:53	:					
ATG	GCT Ala 1	ACC Thr	CCT Pro	TCG Ser	ATG Met 5	ATG Met	CCG Pro	CAG Gln	TGG Trp	TCT Ser 10	TAC Tyr	ATG Met	CAC His	ATC Ile	TCG Ser 15	48
GGC Gly	CAG Gln	GAC Asp	GCC Ala	TCG Ser 20	GAG Glu	TAC Tyr	CTG Leu	AGC Ser	CCC Pro 25	GGG Gly	CTG Leu	GTG Val	CAG Gln	TTT Phe 30	GCC Ala	96
CGC Arg	GCC Ala	ACC Thr	GAG Glu 35	ACG Thr	TAC Tyr	TTC Phe	AGC Ser	CTG Leu 40	AAT Asn	AAC Asn	AAG Lys	TTT Phe	AGA Arg 45	AAC Asn	CCC Pro	144
ACG Thr	GTG Val	GCG Ala 50	CCT Pro	ACG Thr	CAC His	GAC Asp	GTG Val 55	ACC Thr	ACA Thr	GAC Asp	CGG Arg	TCC Ser 60	CAG Gln	CGT Arg	TTG Leu	192
					CCT Pro											. 240
AAG Lys 80	GCG Ala	CGG Arg	TTC Phe	ACC Thr	CTA Leu 85	GCT Ala	GTG Val	GGT Gly	GAT Asp	AAC Asn 90	CGT Arg	GTG Val	CTG Leu	GAC Asp	ATG Met 95	288
GCT Ala	TCC Ser	ACG Thr	TAC Tyr	TTT Phe 100	GAC Asp	ATC Ile	CGC Arg	GGC Gly	GTG Val 105	CTG Leu	GAC Asp	AGG Arg	GGC Gly	CCT Pro 110	ACT Thr	330
TTT Phe	AAG Lys	CCC Pro	TAC Tyr 115	TCT Ser	GGC Gly	ACT Thr	GCC Ala	TAC Tyr 120	AAC Asn	GCC Ala	CTG Leu	GCT Ala	CCC Pro 125	AAG Lys	GGT Gly	384
GCC Ala	CCA Pro	AAT Asn 130	CCT Pro	TGC Cys	GAA Glu	TGG Trp	GAT Asp 135	GAA Glu	GCT Ala	GCT Ala	ACT Thr	GCT Ala 140	CTT Leu	GAA Glu	ATA Ile	432
AAC	CTA	GAA	GAA	GAG	GAC	GAT	GAC	AAC	GAA	GAC	GAA	GTA	GAC	GAG	CAA	480

Asn	Leu 145	Glu	Glu	Glu	Asp	Asp 150	Asp	Asn	Glu	Asp	Glu 155		Asp	Glu	Gln	
GCT Ala 160	GIU	CAG Gln	CAA Gln	AAA Lys	ACT Thr 165	His	GTA Val	TTT Phe	GGG Gly	CAG Gln 170	GCG	CCT Pro	TAT	TCT	GGT Gly 175	528
ATA Ile	AAT Asn	ATT Ile	ACA Thr	AAG Lys 180	GAG Glu	GGT Gly	ATT Ile	CAA Gln	ATA Ile 185	Gly	GTC Val	GAA Glu	GGT Gly	CAA Gln 190	ACA Thr	576
CCT	AAA Lys	TAT Tyr	GCC Ala 195	Asp	AAA Lys	ACA Thr	TTT Phe	CAA Gln 200	Pro	GAA Glu	CCT Pro	CAA Gln	ATA Ile 205	Gly	GAA Glu	624
TCT Ser	CAG Gln	TGG Trp 210	TAC Tyr	GAA Glu	ACT Thr	GAA Glu	ATT Ile 215	Asn	CAT His	GCA Ala	GCT Ala	GGG Gly 220	AGA Arg	GTC Val	CTT Leu	672
AAA Lys	AAG Lys 225	ACT Thr	ACC Thr	CCA Pro	ATG Met	AAA Lys 230	CCA Pro	TGT Cys	TAC	GGT Gly	TCA Ser 235	TAT Tyr	GCA Ala	AAA Lys	CCC Pro	720
240	AAT Asn	Glu	Asn	GIY	G1y 245	Gln	Gly	Ile	Leu	Val 250	Lys	Gln	Gln	Asn	Gly 255	768
AAG Lys	CTA Leu	GAA Glu	AGT Ser	CAA Gln 260	GTG Val	GAA Glu	ATG Met	CAA Gln	TTT Phe 265	TTC Phe	TCA Ser	ACT Thr	ACT Thr	GAG Glu 270	GCG Ala	816
ACC Thr	GCA Ala	GGC Gly	AAT Asn 275	GGT Gly	GAT Asp	AAC Asn	TTG Leu	ACT Thr 280	CCT Pro	AAA Lys	GTG Val	GTA Val	TTG Leu 285	TAC Tyr	AGT Ser	864
GAA Glu	GAT Asp	GTA Val 290	GAT Asp	ATA Ile	GAA Glu	ACC Thr	CCA Pro 295	GAC Asp	ACT Thr	CAT His	ATT Ile	TCT Ser 300	TAC Tyr	ATG Met	CCC Pro	912
ACT Thr	ATT Ile 305	AAG Lys	GAA Glu	GGT Gly	AAC Asn	TCA Ser 310	CGA Arg	GAA Glu	CTA Leu	ATG Met	GGC Gly 315	CAA Gln	CAA Gln	TCT Ser	ATG Met	960
CCC Pro 320	AAC Asn	AGG Arg	CCT Pro	AAT Asn	TAC Tyr 325	ATT Ile	GCT Ala	TTT Phe	AGG Arg	GAC Asp 330	AAT Asn	TTT Phe	ATT Ile	GGT Gly	CTA Leu 335	1008
ATG Met	TAT Tyr	TAC Tyr	AAC Asn	AGC Ser 340	ACG Thr	GGT Gly	AAT Asn	ATG Met	GGT Gly 345	GTT Val	CTG Leu	GCG Ala	GGC Gly	CAA Gln 350	GCA Ala	1056
TCG Ser	CAG Gln	TTG Leu	AAT Asn 355	GCT Ala	GTT Val	GTA Val	GAT Asp	TTG Leu 360	CAA Gln	GAC Asp	AGA Arg	AAC Asn	ACA Thr 365	GAG Glu	CTT Leu	1104
TCA Ser	TAC Tyr	CAG Gln 370	CTT Leu	TTG Leu	CTT Leu	GAT Asp	TCC Ser 375	ATT Ile	GGT Gly	GAT Asp	AGA Arg	ACC Thr 380	AGG Arg	TAC Tyr	TTT Phe	1152
TCT Ser	ATG Met 385	TGG Trp	AAT Asn	CAG Gln	GCT Ala	GTT Val 390	GAC Asp	AGC Ser	TAT Tyr	GAT Asp	CCA Pro 395	GAT Asp	GTT Val	AGA Arg	ATT Ile	1200
ATT	GAA	AAT	CAT	GGA	ACT	GAA	GAT	GAA	CTT	CCA	AAT	TAC	TGC	TTT	CCA	1248

Ile 400	Glu	Asn	His	Gly	Thr 405	Glu	Asp	Glu	Leu	Pro 410	Asn	Tyr	Cys	Phe	Pro 415		
													AAA Lys				1296
													TTT Phe 445				1344
													ATC Ile				1392
													GCG Ala				1440
													ATT Ile				1488
													GCT Ala				1536
													CTT Leu 525				1584
ATG Met	GAC Asp	AAC Asn 530	GTC Val	AAC Asn	CCA Pro	TTT Phe	AAC Asn 535	CAC His	CAC His	CGC Arg	AAT Asn	GCT Ala 540	GGC Gly	CTG Leu	CGC Arg	•	1632
													TTC Phe				1680
													CTC Leu				1728
			Thr	Tyr		Trp	Asn	Phe	Arg	Lys	Asp	Val	AAC Asn				1776
													GCC Ala 605				1824
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													ACC Thr				1920
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	GCT Ala								2064
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	GCC Ala								2208
	CTT Leu								2256
	TAC Tyr								2304
	ATG Met								2352
	AGC Ser 785								2400
	AGC Ser								2448
•	GGC Gly								2496
	CCC Pro								2544
	CTT Leu								2592
	TGC Cys 865								2640
	ATG Met								2688
	GCC Ala								2736

	ACC Thr															2784
	CGG Arg															2832
	TCG Ser 945							AA								2858
(2)	INFO	RMAT	NOI	FOR	SEQ	ID N	10:54	1:								
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	(ii)	MOI	LECUI	LE TY	PE:	prot	ein									
	(xi)	SEÇ	QUENC	CE DI	ESCR	PTIC	ом: s	SEQ :	ID NO	54:	:					
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Ala	Thr	Glu 35	Thr	Tyr	Phe	Ser	Leu 40	Asn	Asn	Lys	Phe	Arg 45	Asn	Pro	Thr	
Val	Ala 50	Pro	Thr	His	Asp	Val 55	Thr	Thr	Asp	Arg	Ser 60	Gln	Arg	Leu	Thr	
Leu 65	Arg	Phe	Ile	Pro	Val 70	Asp	Arg	Glu	Asp	Thr 75	Ala	Tyr	Ser	Tyr	Lys 80	
Ala	Arg	Phe	Thr	Leu 85	Ala	Val	Gly	Asp	Asn 90	Arg	Val	Leu	Asp	Met 95	Ala	
Ser	Thr	Tyr	Phe 100	Asp	Ile	Arg	Gly	Val 105	Leu	Asp	Arg	Gly	Pro 110	Thr	Phe	
Lys	Pro	Tyr 115	Ser	Gly	Thr	Ala	Tyr 120	Asn	Ala	Leu	Ala	Pro 125	Lys	Gly	Ala	
Pro	Asn 130	Pro	Суз	Glu	Trp	Asp 135	Glu	Ala	Ala	Thr	Ala 140	Leu	Glu	Ile	Asn	
Leu 145	Glu	Glu	Glu	Asp	Asp 150	Asp	Asn	Glu	Asp	Glu 155	Val	Asp	Glu	Gln	Ala 160	
Glu	Gln	Gln	Lys	Thr 165	His	Val	Phe	Gly	Gln 170	Ala	Pro	Tyr	Ser	Gly 175	Ile	
Asn	Ile	Thr	Lys 180	Glu	Gly	Ile	Gln	Ile 185	Gly	Val	Glu	Gly	Gln 190	Thr	Pro	
Lys	Tyr	Ala 195	Asp	Lys	Thr	Phe	Gln 200	Pro	Glu	Pro	Gln	11e 205	Gly	Glu	Ser	
Gln	Trp 210	Tyr	Glu	Thr	Glu	Ile 215	Asn	His	Ala	Ala	Gly 220	Arg	Val	Leu	Lys	

Lys 225	Thr	Thr	Pro	Met	Lys 230	Pro	Cys	Tyr	Gly	Ser 235	Tyr	Λla	Lys	Pro	Thr 240
Asn	Glu	Asn	Gly	Gly 245	Gln	Gly	Ile	Leu	Val 250	Lys	Gln	Gln	Asn	Gly 255	Lys
Leu	Glu	Ser	Gln 260	Val	Glu	Met	Gln	Phe 265	Phe	Ser	Thr	Thr	Glu 270	Ala	Thr
Ala	Gly	Asn 275	Gly	Asp	Asn	Leu	Thr 280	Pro	Lys	Val	Val	Leu 285	Tyr	Ser	Glu
Asp	Val 290	Asp	Ile	Glu	Thr	Pro 295	Asp	Thr	His	Ile	Ser 300	Tyr	Met	Pro	Thr
11e 305	Lys	Glu	Gly	Asn	Ser 310	Arg	Glu	Leu	Met	Gly 315	Gln	Gln	Ser	Met	Pro 320
Asn	Arg	Pro	Asn	Tyr 325	Ile	Ala	Phe	Arg	Asp 330	Asn	Phe	Ile	Gly	Leu 335	Met
Tyr	Tyr	Asn	Ser 340	Thr	Gly	Asn	Met	Gly 345	Val	Leu	Ala	Gly	Gln 350	Ala	Ser
Gln	Leu	Asn 355	Ala	Val	Val	Asp	Leu 360	Gln	Asp	Arg	Asn	Thr 365	Glu	Leu	Ser
Tyr	Gln 370	Leu	Leu	Leu	Asp	Ser 375	Ile	Gly	Asp	Arg	Thr 380	Arg	Tyr	Phe	Ser
Met 385	Trp	Asn	Gln	Ala	Val 390	Asp	Ser	Tyr	Asp	Pro 395	Asp	Val	Arg	Ile	Ile 400
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Gly	Gly	Val	Ile 420	Asn	Thr	Glu	Thr	Leu 425	Thr	Lys	Val	Lys	Pro 430	Lys	Thr
Gly	Gln	Glu 435	Asn	Gly	Trp	Glu	Lys 440	Asp	Ala	Thr	Glu	Phe 445	Ser	Asp	Lys
Asn	Glu 450	Ile	Arg	Val	Gly	Asn 455	Asn	Phe	Ala	Met	Glu 460	Ile	Asn	Leu	Asn
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Val	Asp	Cys 515	Tyr	Ile	Asn	Leu	Gly 520	Ala	Arg	Trp	Ser	Leu 525	Asp	Tyr	Met
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Arg 545	Ser	Met	Leu	Leu	Gly 550	Asn	Gly	Arg	Tyr	Val 555	Pro	Phe	His	Ile	Gln 560

Val	Pro	Gln	Lys	Phe 565	Phe	Ala	Ile	Lys	Asn 570	Leu	Leu	Leu	Leu	Pro 575	Gly
Ser	Tyr	Thr	Tyr 580	Glu	Trp	Asn	Phe	Arg 585	Lys	Asp	Val	Asn	Met 590	Val	Leu
Gln	Ser	Ser 595	Leu	Gly	Asn	Asp	Leu 600	Arg	Val	Asp	Gly	Ala 605	Ser	Ile	Lys
Phe	Asp 610	Ser	Ile	Cys	Leu	Tyr 615	Ala	Thr	Phe	Phe	Pro 620	Met	Ala	His	Asn
Thr 625	Ala	Ser	Thr	Leu	Glu 630	Ala	Met	Leu	Arg	Asn 635	Asp	Thr	Asn	Asp	G1n 640
Ser	Phe	Asn	Asp	Tyr 645	Leu	Ser	Ala	Ala	Asn 650	Met	Leu	Tyr	Pro	Ile. 655	Pro
Ala	Asn	Ala	Thr 660	Asn	Val	Pro	Ile	Ser 665	Ile	Pro	Ser	Arg	Asn 670	Trp	Ala
Ala	Phe	Arg 675	Gly	Trp	Ala	Phe	Thr 680	Arg	Leu	Lys	Thr	Lys 685	Glu	Thr	Pro
Ser	Leu 690	Gly	Ser	Gly	Tyr	Asp 695	Pro	Tyr	Tyr	Thr	Tyr 700	Ser	Gly	Ser	Ile
Pro 705	Tyr	Leu	Asp	Gly	Thr 710	Phe	Tyr	Leu	Asn	His 715	Thr	Phe	Lys	Lys	Val 720
Ala	Ile	Thr	Phe	Asp 725	Ser	Ser	Val	Ser	Trp 730	Pro	Gly	Asņ	Asp	Arg 735	Leu
Leu	Thr	Pro	Asn 740	Glu	Phe	Glu	Ile	Lys 745	Arg	Ser	Val	Asp	Gly 750	Glu	Gly
Tyr	Asn	Val 755	Ala	Gln	Cys	Asn	Met 760	Thr	Lys	Asp	Trp	Phe 765	Leu	Val	Gln
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Ser 785	Tyr	Lys	Asp	Arg	Met 790	Tyr	Ser	Phe	Phe	Arg 795	Asn	Phe	Gln	Pro	Met 800
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Gly	Ile	Leu	His 820	Gln	His	Asn	Asn	Ser 825	Gly	Phe	Val	Gly	Tyr 830	Leu	Ala
Pro	Thr	Met 835	Arg	Glu	Gly	Gln	Ala 840	Tyr	Pro	Ala	Asn	Phe 845	Pro	Tyr	Pro
Leu	Ile 850	Gly	Lys	Thr	Ala	Val 855	Asp	Ser	Ile	Thr	Gln 860	Lys	ŗ'ns	Phe ,	Leu
Cys 865	Asp	Arg	Thr	Leu	Trp 870	Arg	Ile	Pro	Phe	Ser 875	Ser	Asn	Phe	Met	Ser 880
Met	Gly	Ala	Leu	Thr 885	Asp	Leu	Gly	Gln	Asn 890	Leu	Leu	Tyr	Ala	Asn 895	Ser

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									102							
Ala	His-	Ala	Leu 900	Asp	Met	Thr	Phe	Glu 905	Val	Asp	Pro	Met	Asp 910	Glu	Pro	
Thr	Leu	Leu 915	Tyr	Val	Leu	Phe	Glu 920	Val	Phe	Asp	Val	Val 925	Arg	Val	His	
Arg	Pro 930	His	Arg	Gly	Val	Ile 935	Glu	Thr	Val	Tyr	Leu 940	Arg	Thr	Pro	Phe	
Ser 945	Ala	Gly	Asn	Ala	Gln 950	His			_							
(2)	INFO	RMAT	ION	FOR	SEQ	ID N	10:55	; :								
		(A (B (C	A) LE B) TY C) ST D) TO	NGTH PE: RAND	IARACI: 98 nucl DEDNE DGY:	bas eic ISS: line	se pa acid doub ar	airs i ole	.c ac	eid						
	(xi)	SEÇ	UENC	E DE	SCRI	PTIC	on: s	SEQ 1	D NO	55:						
GAA Glu 1	CTC Leu	GGA Gly	GGT Gly	GGA Gly 5	GGT Gly	GGA Gly	ACT Thr	AGT Ser	TTT Phe 10	GGA Gly	CGC Arg	GGA Gly	GAC Asp	ATT Ile 15	CGC Arg	48
AAT Asn	TAAA	GTAC	TG G	SATTO	ATGA	C TC	CTAGA	CTT	ATI	AAGO	SATC	CAAI	AAA			98
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	(i)	(A (B	() I	ENGI PE:	IARAC 'H: 1 amir GY:	7 am	ino id		ls							
	(ii)	MOL	ECUL	E TY	PE:	pept	ide									
	(xi)	SEQ	UENC	E DE	SCRI	PTIC	N: S	EQ I	D NC	:56:						

Glu Leu Gly Gly Gly Gly Gly Thr Ser Phe Gly Arg Gly Asp Ile Arg
1 10 15

Asn

WHAT IS CLAIMED IS:

- 1. A chimeric adenovirus coat protein comprising a nonnative amino acid sequence, wherein said chimeric adenovirus coat protein has a decreased ability or inability to be recognized by a neutralizing antibody directed against the wild-type adenovirus coat protein.
- 2. The chimeric adenovirus coat protein of claim 1, wherein said nonnative amino acid sequence comprises a deletion, insertion, or a replacement of a region of from about 1 to about 750 amino acids of said wild-type adenovirus coat protein.
- 3. The chimeric adenovirus coat protein of claim 1 or 2, wherein said nonnative amino acid sequence comprises a plurality of deletions, insertions, and/or replacements.
- 4. The chimeric adenovirus coat protein of any of claims 1-3, wherein said coat protein is a chimeric adenovirus hexon protein.
- 5. The chimeric adenovirus coat protein of claim 4, wherein said region deleted or replaced comprises a hypervariable region in either the 11 loop or the 12 loop.
- 6. The chimeric adenovirus coat protein of claim 5, wherein said hypervariable region is selected from the group consisting of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, and HVR7.
- 7. The chimeric adenovirus coat protein of any of claims 1-6, comprising a sequence selected from the group consisting of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID

- NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48.
- 8. The chimeric adenovirus coat protein of any of claims 1-7, wherein said nonnative amino acid sequence comprises a spacer of about 1 to about 750 amino acids.
- 9. The chimeric coat adenovirus coat protein of claim 8, wherein said spacer comprises the sequence of SEQ ID NO:50.
- 10. The chimeric adenovirus coat protein of any of claims 1-9, comprising an amino acid sequence of a coat protein of another serotype of adenovirus.
- 11. The chimeric adenovirus coat protein of claim 10, wherein said coat protein of another serotype is a hexon protein.
- 12. An isolated or purified nucleic acid that encodes the chimeric adenovirus coat protein of any of claims 1-11.
- 13. The isolated or purified nucleic acid of claim 12 comprising a sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, and SEQ ID NO:47.

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- 14. The isolated or purified nucleic acid of claim12 or 13 comprising SEQ ID NO:49.
- 15. An adenoviral vector that comprises the chimeric adenovirus coat protein of any of claims 1-11.
- 16. A method of genetically modifying a cell which comprises contacting said cell with the adenoviral vector of claim 15.
- 17. A host cell that comprises the chimeric adenovirus coat protein of any of claims 1-11.
- 18. A method of constructing an adenoviral vector that has a decreased ability or inability to be recognized by a neutralizing antibody directed against wild-type adenovirus coat protein, which method comprises obtaining an adenoviral vector comprising a wild-type adenovirus coat protein and replacing said wild-type adenovirus coat protein with the chimeric adenovirus coat protein of any of claims 1-11.

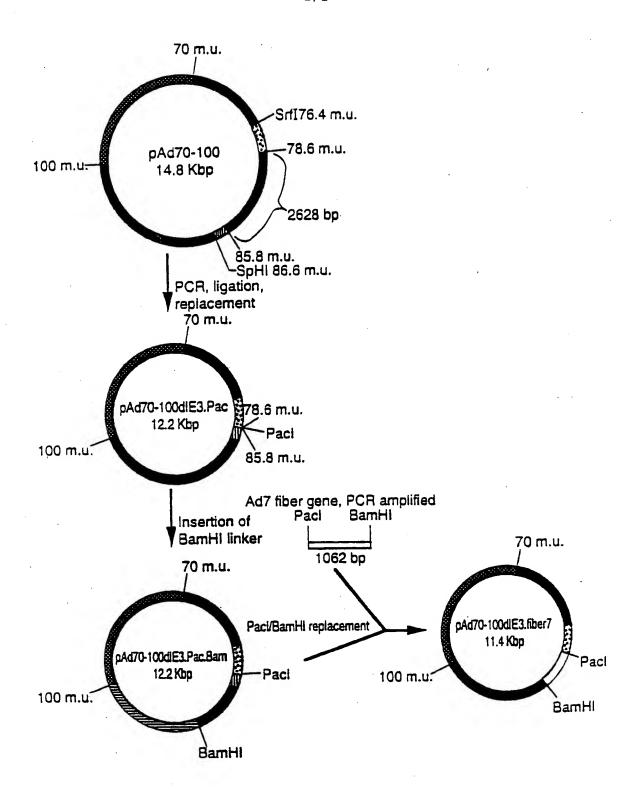


FIG. 1

SUBSTITUTE SHEET (RULE 26)

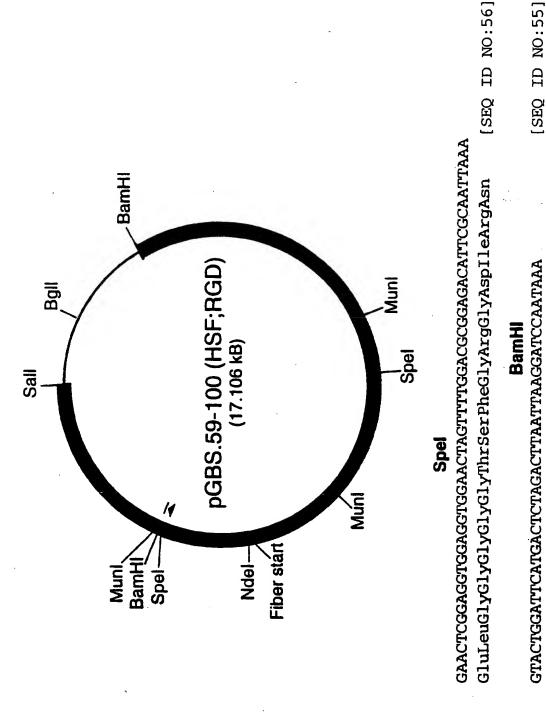


FIG. 2

INTERNATIONAL SEARCH REPORT

im. .ttonal Application No PCT/US 98/05033

A. CLASS	SIFICATION OF SUBJECT MATTER	·	
IPC 6	C12N15/86 C07K14/075 C12N15,	/34 C12N5/10	
According t	to international Patent Classification (IPC) or to both national classifi	cation and IPC	
1	SEARCHED	nasion and it o	
Minimum d	ocumentation searched (classification system followed by classification	tion symbols)	
IPC 6	C12N C07K		
Documenta	ation searched other than minimum documentation to the extent that	such documents are included in the fields se	parched
			_
Electronic o	data base consulted during the international search (name of data b	ase and, where practical, search terms used	
		, , , , , , , , , , , , , , , , , , , ,	,
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C. DOCUM	ENTS CONSIDERED TO BE RELEVANT		
Category 3	Citation of document, with indication, where appropriate, of the re	Nevent nassages	
	, , , , , , , , , , , , , , , , , , , ,	Total passages	Relevant to claim No.
A	WO 96 26281 A (GENVEC, INC.) 29	August	
,,	1996	August	1-18
	see page 5, line 7 - line 23		
	see page 6, line 30 - line 37	•	
Α	COMPTON 1 ST AL HE		
^	CROMPTON J ET AL.: "Expression forein epitope on the surface of	of a	1-18
	adenovirus hexon"	tne	
	JOURNAL OF GENERAL VIROLOGY,		
	vol. 75, no. 1, January 1994. RE	ADING GB.	
	pages 133-139, XP002071015		
	cited in the application	_	
	see table 1		
ĺ		-/ 	i
X Furth	her documents are listed in the continuation of box C.	Patent family members are listed in	n annex.
* Special cat	egories of cited documents :	"T" later document published after the inter	national filling date
"A" docume	nt defining the general state of the art which is not ered to be of particular relevance	or priority date and not in conflict with cited to understand the principle or the	the application but
"E" earlier d	ocument but published on or after the international	invention	
niing ca	ate If which may throw doubts on priority claim(s) or	"X" document of particular relevance; the c cannot be considered novel or cannot	be considered to
Municu a	s cited to establish the publication date of another or other special reason (as specified)	involve an inventive step when the document of particular relevance; the c	
	nt referring to an oral disclosure, use, exhibition or	cannot be considered to involve an im- document is combined with one or mo	rentive step when the
The documen	nt published prior to the international filling date but	ments, such combination being obviou in the art.	is to a person skilled
aterun	an the priority date claimed	"&" document member of the same patent i	amily
Date of the a	ctual completion of theinternational search	Date of mailing of the international season	rch report
a	July 1998	07/07/1000	
		27/07/1998	
Name and m	atiling address of the ISA	Authorized officer	
	European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Rijswijk		
	Tel. (+31-70) 340-2040, Tx. 31 651 epo ni. Fax: (+31-70) 340-3016	Cupido, M	

INTERNATIONAL SEARCH REPORT

In. atlanel Application No
PCT/US 98/05033

		PCT/US 98/05033
	etion) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category "	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	CRAWFORD-MIKSZA L AND SCHNURR P: "Analysis of 15 adenovirus hexon proteins reveal the location and structure of seven hypervariable regions containing serotype-specific residues" JOURNAL OF VIROLOGY, vol. 70, no. 3, March 1996, AMERICAN SOCIETY FOR MICROBIOLOGY US, pages 1836-1844, XP002071016 cited in the application	1-18
Α	MASTRANGELI A ET AL.: ""Sero-switch" Adenovirus-mediated in vivo gene transfer: Circumvention of anti-adenovirus humoral immune defenses against repeat adenovirus vector administration by changing the adenovirus serotype" HUMAN GENE THERAPY, vol. 7, no. 1, 1 January 1996, pages 79-87, XP000653452 cited in the application	1-18
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Information on patent family members

In ational Application No PCT/US 98/05033

Patent document cited in search repor	t	Publication date		Patent family member(s)	Publication date
WO 9626281	Α	29-08-1996	US AU CA EP	5770442 A 4980496 A 2213343 A 0811069 A	23-06-1998 11-09-1996 29-08-1996 10-12-1997

Form PCT/ISA/210 (paters family annex) (July 1992)